Asian Food Science Journal



2(2): 1-9, 2018; Article no.AFSJ.40555

## Biochemical and Nutritional Parameters from Flour of Ackee Blighia sapida (Sapindaceae) Seeds

Titiana Mahugnon Aguemon<sup>1</sup>, Yolande Dogoré Digbeu<sup>2</sup>, Jacques Gnanwa Mankambou<sup>3</sup>, Edmond Ahipo Dué<sup>1\*</sup> and Lucien Patrice Kouamé<sup>1</sup>

<sup>1</sup>Biochemistry and Food Technology Laboratory, Nangui Abrogoua University, 02 BP 801 Abidjan 02, Côte d'Ivoire.

<sup>2</sup>Laboratory of Food Safety, Nangui Abrogoua University, 02 BP 801 Abidjan 02, Côte d'Ivoire. <sup>3</sup>Laboratory of Agrovalorisation, Jean Lorougnon Guede University, BP 150, Daloa, Côte d'Ivoire.

#### Authors' contributions

This work was carried out in collaboration between all authors. Author EAD designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors YDD and TMA managed the analyses of the study. Author JGM managed the literature searches. All authors read and approved the final manuscript.

#### Article Information

DOI: 10.9734/AFSJ/2018/40555 <u>Editor(s):</u> (1) Surapong Pinitglang, Assistant Professor, Department of Food Business Management, School of Science and Technology, University of the Thai Chamber of Commerce, Bangkok, Thailand. <u>Reviewers:</u> (1) Abdullahi Abdulkadir Imam, Bayero University Kano, Nigeria. (2) Barnabas Kayaltos, Burkina Faso. (3) O. T. Talabi, Babcock University, Nigeria. Complete Peer review History: <u>http://www.sciencedomain.org/review-history/24489</u>

> Received 8<sup>th</sup> February 2018 Accepted 17<sup>th</sup> April 2018 Published 7<sup>th</sup> May 2018

**Original Research Article** 

### ABSTRACT

The limited information on the biochemical and nutritional benefits of edible ackee (*Blighia sapida*) seeds makes it underutilized in West Africa. This study was to investigate the nutrient content and antinutritional factors using standard analytical methods. The results revealed that ackee (*B. sapida*) seeds contain some percentage flavonoid ( $0.134 \pm 0.08 \text{ g}/100 \text{ g}$ ), carotenoid ( $0.005 \pm 0.01 \text{ g}/100 \text{ g}$ ), polyphenol ( $0.851 \pm 0.40 \text{ g}/100 \text{ g}$ ), total sugar ( $67.98 \pm 0.27 \text{ g}/100 \text{ g}$ ), reducing sugar ( $86.93 \pm 0.73 \text{ g}/100 \text{ g}$ ), vitamin C ( $0.019 \pm 0.49 \text{ g}/100 \text{ g}$ ), moisture contents ( $6.22 \pm 0.04 \text{ g}/100 \text{ g}$ ), crude fibre ( $13.84 \pm 0.08 \text{ g}/100 \text{ g}$ ), ash ( $2.68 \pm 0.09 \text{ g}/100 \text{ g}$ ), crude protein ( $7.83 \pm 0.04 \text{ g}/100 \text{ g}$ ) and carbohydrate ( $75.77 \pm 0.08 \text{ g}/100 \text{ g}$ ). The antinutritional composition which include alkaloids, oxalate, phytate and tannin were  $0.23 \pm 0.02$ ,  $391.87 \pm 0.07$ ,  $415.09 \pm 0.70$  and  $795.00 \pm 0.37$ 

mg/100 g respectively. These results indicated that ackee seeds are good source of nutritional compounds. However, they had considerable antinutritional factors. Thus, it would be better to treat them by thermal processing methods before using them for application in food systems to maintain food quality.

Keywords: Ackee (Blighia sapida); seeds; biochemical; nutritional; antinutritional factors.

#### **1. INTRODUCTION**

The ackee (*Blighia sapida* Köenig) belongs to the *Sapindaceae* and is a native plant of West Africa that was introduced to Jamaica in the 18th century. The fruit is a three-celled fleshy capsule containing three valves and three glossy black seeds. When ripe, the fruit splits longitudinally into three sections to reveal the seeds and the thick yellow flesh of the arils, which form the edible portion of the fruit and has a nutty flavor, [1,2].

The ripe arils of the ackee fruit, yellow to cream coloured, are nutty-flavored and edible [3]. The arils are the major component of the Jamaican national dish; ackee and saltfish [4]. The ripe fruit arils are eaten fresh, dried, fried, roasted or made into sauce or soup in some parts of West Africa [5]. Thus, ackee arils have been reported to have comparable proximate composition to many known legumes and oil seeds [6,7,3]. Various parts from ackee as the roots, bark, leaves, capsules and seeds were identified in the treatment of 22 diseases in Ashanti region of Ghana. [8]. However, ackee seeds have little commercial and nutritional significance in the West African sub-region.

Substantial scientific knowledge on the nutritional beneficial constituents from ackee seeds could ensure the development of more efficient ways to convert the fruit into useful products with improved commercial value. This study, therefore, aims to investigate the biochemical contents, nutritional and antinutritional factors of ackee seeds from Côte d'Ivoire that may be useful in its application as nutritional, industrial and pharmaceutical base.

#### 2. MATERIALS AND METHODS

#### 2.1 Raw Materials

The biological material used for this study is from ackee (*Blighia sapida*) seeds (Fig. 1). The ackee is a spontaneous plant which period of availability is may-June with local names, Kaa (*Center of* Côte d'Ivoire), Finzan (north of Côte

d'Ivoire), Atuanbi (south of Côte d'Ivoire); and Goihien (West of Côte d'Ivoire). The ackee seeds, predominant in north region, were harvested at Lakota (Center-West of Côte d'Ivoire (West Africa). They were immediately transported to the Laboratory of Biocatalysis and Bioprocessing of University Nangui Abrogoua (Abidjan, Côte d'Ivoire) and stored under prevailing tropical ambient conditions before the preparation of flours from raw.



#### Fig. 1. Blighia sapida seeds

All chemicals and reagents used were of analytical grade and purchased from Sigma Chemical Company (USA).

#### 2.2 Sample Preparation

The ackee (*Blighia sapida*) seeds were thoroughly sorted to remove bad ones from the lot. The retained seeds were washed with clean water to eliminate adhering dirt and extraneous materials, then they were dried in an oven at  $65^{\circ}$ C for 48 hours. After these steps, the dried seeds were ground into powder, sieved with 250 µm mesh sieve and stored in bottles in an oven at  $55^{\circ}$ C for different analysis (Fig. 2).

### 2.3 Physico-chemical Composition Analysis

Dry matters were determined by drying in an oven at  $105 \pm 2^{\circ}$ C during 24 h to constant weight

as described by Association of Official Analytical Chemists (AOAC) [9]. The total ash contents were determined by incinerating flour (10 g) in a furnace at 550 ± 15°C for 12 hours, then weighing the residue after cooling to room temperature in a desiccator (AOAC) [9]. Crude protein was calculated from nitrogen (Nx6.25) obtained using the Kjeldahl method by AOAC [9]. The carbohydrate contents were determined by deference that is by deducting the mean values of other parameters that were determined from 100. Therefore % carbohydrate = 100 - (% moisture +% crude protein + % crude fat + crude fibre + % ash) [10,11]. The crude fibre contents were determined according to standard method (AOAC, [9]. Method described by Dubois et al. [12] was used to determine total sugars while reducing sugars were analyzed according to the method of Bernfeld [13], using 3.5 dinitrosalycilic acids (DNS).



## Fig. 2. Flow diagram of the process for producing *B.sapida* seed flour

#### 2.4 Biochemical Analysis

Vitamin C (ascorbic acid) content was determined by the method of Pongracz et al. [14] and Barros et al. [15]. Total phenolics compound contents were determined as described by Hanson et al. [16] from the methanol extracts using Folin-Ciocalteu reagent [17]. carotenoid content was determined according to the method of Rodriguez-Amaya [18].

#### 2.5 Antinutrients Analysis

Oxalate content was determined by the method of Day and Underwood [19]. Tannin content,

flavonoid content and phytate content were estimated by the spectrophotometric method described respectively by Bainbridge et al. [20], Meda et al. [21] and INRA [22]. Alkaloid content was estimated by the filtration method of Harbone [23].

Each of the samples from ackee (*Blighia sapida*) flour was analyzed in triplicate for their biochemical composition, physico-chemical and antinutrients properties.

## 2.6 Statistical Analyses

Statistical analyses were carried out in triplicate. The results were processed by the software STATISTICA 7 (Stat soft Inc, Tulsa-USA, Headquarters). Thus, results were expressed as means ± standard deviation. The statistical differences among the means of data were calculated using one-way analysis of variance (ANOVA) and Duncan's Multiple Range Test (DMRT). Differences at P<0.05 were considered significant.

#### **3. RESULTS AND DISCUSSION**

## **3.1 Biochemical Content**

The biochemical composition of ackee (*Blighia* sapida) seed is shown in Table 1. The results show high content of reducing sugar and total sugar ( $86.93 \pm 0.73$  g/100 g and  $67.98 \pm 0.27$  g/100 g) respectively. But they show low content of vitamin C (0.019 g/100 g), carotenoid (0.005 ± 0.01 g/100 g), polyphenol ( $0.851\pm 0.40$  g/100 g) and flavonoid ( $0.134 \pm 0.08$  g/100 g),

Flavonoid is polyphenolic compound that is ubiquitous in nature, comprising a number of hydroxyl groups attached to aromatic ring structures that determine its antioxidative compounds properties. The exhibit а diphenylpropane (C6-C3-C6) skeleton and multiple sub-groups. The quantity of this compound is important in justifying the antioxidative properties of the seeds. The value found in this study is higher compared to those reported by Buratto et al. [24] which have analyzed the content of total flavonoids in Brazil nuts, they have obtained 0.34 g EQ/kg of dry sample. This value is also higher than those found in chia seed extracts. In a study by Lin and Tang [25] where it was evaluated the content of total flavonoids in vegetables, the values were 0.075 g EQ/kg for green peppers, 0.041g EQ/kg for yellow pepper and 0.306 g EQ/kg for white onion.

Biochemical		Nutritional	
Parameters	Values (g/100 g DM)	Parameters	Values (g/100 g DM)
Flavonoid	0.134 ± 0.08	Moisture	6.22 ± 0.04
Carotenoid	0.005 ± 0.01	Fiber	13.84 ± 0.08
Total sugar	67.98 ± 0.27	Ash	2.68 ± 0.09
Reducing sugar	86.93 ± 0.73	Protein	7.83 ± 0.04
Vitamin C	0.019 ± 0.49	Carbohydrates	75.77 ± 0.08
Polyphenol	0.851 ± 0.40		

Table 1. Biochemical and nutritional contents from flour of ackee B. sapida seed

Statistical analyses were carried out in triplicate

Phenolic compound is natural antioxidant found generally in several fruits of both trees and compound has maximum cereals. The concentration at the superficial layers of the kernel, which set-up the branch Proestos et al. [26]. Phenolics have been found as strong antioxidants towards hindering the influence of free radicals and ROS, which is the basis of several chronic human infections [27]. The presence of significant phenolic compound from ackee Blighia sapida seeds will encourage the utilization of the seeds for many purposes. The phenolics and flavonoids are important indicators of antioxidant capacity of ackee Blighia sapida seeds. These compounds play vital function towards preventing diseases and sustain a state of well being.

The roles of carotenoids in seed are less clear than in other tissues, but are emerging. Carotenoid production in the seed is important for ABA production and seed dormancy [28]. Furthermore, carotenoids contribute to the antioxidant system in seeds, which functions to limit free radical-induced membrane deterioration and seed ageing [29,30].

Carotenoids are the precursors of vitamin A and similar compounds. ß- carotene is one of most commonly known carotenoids which is a potent antioxidant as well as a dietary factor for growth. It is a precursor of vitamin A that has important role in vision, as the prosthetic group of the light sensitive proteins in retina, and a major role in the regulation of gene expression and tissue differentiation [31]. Deficiency of vitamin A is a major public health problem around the world. The prevention of vitamin A deficiency is one of the three micronutrient priorities of the World Health Organization (WHO), others are iron and iodine.

Vitamin C, or ascorbic acid, is a water soluble antioxidant that plays a vital role in protecting the body from infection and disease. It is not synthesised by the human body and therefore must be acquired from dietary sources primarily fruits and vegetables. Losses of vitamin C are mostly as a result of leaching into the processing water, thermal destruction and oxidation [32]. It is to be noted that the requirement of vitamin C increases durina pregnancy, lactation, adolescence, hyperthyroidism, infection and after surgery [33]. Maintenance of daily dietary intake of vitamin C leads to the prevention of scurvy which is the deficiency disease state of vitamin C. The fairly high ascorbic acid value from flour of ackee seed gives an indication that the seeds may be good source of ascorbic acid when compared to citrus fruits [34].

#### **3.2 Nutritional Content**

The result revealed that the moisture content from ackee B. sapida seed was 6.22 ± 0.04 g/100 g. This value is similar to 5.24 ± 0.05 g/100 g and lower than 11.0 g/100 g reported by Ogungbenle [35]. It is lower than the moisture content of different varieties of date palm as 14.81 ± 0.396 (Dora), 9.90 ± 0.042 (Dhaki), 12.3 ± 0.242 (Karbaline) respectively was reported by Fagir et al. [36]. The result of Rehman et al. [37] also shows higher content of 17.70 ± 0.03 g/100 g for hard date palm. It noticed that these differences may be due to the location, time, environments, longitivity and maturity of the sample used for the analysis. According to Rodrigues, [38] the great variability in the physical and chemical characteristics of plant can be attributed to many factors, including the region where the plant was grown, climatic differences, fertility, soil pH and annual rainfall. The low moisture content as saw in the sample is an evidence that the ackee Blighia sapida seed specimen may not be more inclined to decay, since nourishments with high dampness substance are more inclined to perishability. [39]. Moisture content of any food is an index of its water activity and is used as a measure of stability and susceptibility to microbial contamination [40]. It might be profitable in perspective of the specimen timeframe of realistic usability.

Crude fibre decreases the absorption of cholesterol from the gut in addition to delaving the digestion and conversion of starch to simple sugars, an important factor in the management of diabetes [41]. Dietary fiber serves as a useful tool in the control of oxidative processes in food products and as functional food ingredient [42]. The crude fiber content from flour of ackee seed was 13.84 ± 0.08 g/100 g. This value is higher than 4.34 ± 0.03 g/100 g and 4.00 ± 0.02 g/100 g obtained by Ogungbenle [35] and Rehman et al. [37]. This result is also higher than 9.4 ± 0.10 g/100 g reported by Gamal et al. [43]. It reported that fiber has some physiological effect in the gastrointestinal track [44] and low fiber in diet is undesirable as it may cause constipation. The high content of ackee seed fiber can enhance satiety, regulate intestinal transit, reduce energy consumption and promote weight loss in users [45].

Ash represents the mineral matter left after food material is burnt in oxygen [46]. It is used as a tool to measure the mineral content in any sample [46]. The ackee B. sapida had moderately high value of ash (2.68 ± 0.09 g/100 g), which indicates that the seed can serve as a viable tool for nutritional evaluation [47]. This result is within the acceptable ash content mean values of legumes of 2.4 to 5.0 % recommended by FAO [48]. The result of the ash content in the sample is a suggestion of a low deposit of mineral elements in the samples compare to the recommended values by the FAO. This may indicate that ackee seed would likely contain very high qualities essential minerals. Since ash content is an index to evaluate and grade the nutritive quality of foods [49].

The crude protein content from flour of ackee *Blighia sapida* seed (7.83  $\pm$  0.04 g/100 g) was lower when compared to oven dried ackee arils (11.67  $\pm$  0.37 g/100 g) [50] and to certain common legume *Phaseolus vulgaris* (20.9 g/100 g); *Lenus culinaris* (20.6 g/100 g) and *Cicer arietinum* (18.5 g/100 g) [51]. It is also lower than that of *Canavalia gladiate* (27.48 g/100 g) and *Canavalia ensiformis* (30.62 g/100 g) [52]. It is generally known that any plant that provides more than 12% of their caloric value from protein is considered to be a good source of protein. Thus ackee *B. sapida* is not a good source of protein.

The carbohydrate content  $(75.77 \pm 0.08 \text{ g}/100 \text{ g})$ from flour of ackee seed is higher than that of like Bambara most legume groundnut (65 g/100 g), broad bean (56.9 g/100 g), chick peas (60.9 g/100 g) etc [53]. Its value (75.77 ± 0.08 g/100 g) was also higher than that of soya beans (32 g/100 g), groundnut (21.0 g/100 g) etc. [53]. But this value is slightly lower than 80.67± 0.05 g/100 g obtained by Ogungbenle [35]. It reported that carbohydrate provides energy to the cells in the body, particularly the brain, which is the only carbohydrate-dependent organ in the body [46]. It is necessary for maintenance of the plasma level, it spares the body protein from being easily digested and helps to prevent the using up of protein. The fairly high carbohydrate content found in flour of ackee seeds suggests high caloric value.

### 3.3 Determination of Antinutritional Factors

The nutritional importance of a given food depends on the nutrients and anti-nutritional constituents of the food [54]. The result of the anti-nutritional factors from flour of ackee *B. sapida* seed is presented in Table 2.

# Table 2. Antinutritional factors from flour of ackee B. sapida seed

Parameters	Values (mg/100 g DM)	
Alkaloid	230 ± 0.02	
Tannin	795.00 ± 0.37	
Oxalate	391.87 ± 0.07	
Phytate	415.09 ± 0.70	
Otatistical analyse	an increase and and and in the line to	

Statistical analyses were carried out in triplicate

The antinutritional composition of flour from ackee seeds (Alkaloid, tannin, oxalate and phytate content) was very high. The alkaloid content only was low with  $230 \pm 0.02$  mg/100 g, however oxalate, phytate and tannin content was high with  $391.87 \pm 0.07$ ,  $415.09 \pm 0.70$  and  $795.00 \pm 0.37$  mg/100g respectively.

Oxalate have been shown to have negative impact on accessibility of mineral which will prompt assimilation of fundamental minerals in body particularly calcium by framing insoluble salts [55]. According to Savage, [56] oxalate and its contents have deleterious effects on human nutrition and health, mainly by decreasing calcium absorption and aiding the formation of kidney stones. The formation of oxalate crystal is said to take place in digestive tract, [57], Nkafamiya et al. [58] have evaluated some oxalate content from seeds of the fruits of some wild plants. *Gemlina arborea* is one of them. They observed  $9.16 \pm 0.13 \text{ mg}/100 \text{ g}$  oxalate in seeds of *G. arborea* and  $50 \pm 0.06 \text{ mg}/100 \text{ g}$  in fruits. The calcium oxalate which is insoluble may cause kidney stone, [59] was also reported by Bello et al., [60].

Phytic acid is considered an antinutrient, as it interferes with the daily activities of human body like digestion and protein breakdown [61]. In this study concentration of phytic acid was found to be 415.09 ± 0.70 mg/100 g. Aberoumand [62] carried out the screening of phytochemical and anti-nutrients compounds of eight food plants sources. The phytate content of these plants ranged from 248.0 mg/100 g to 823.6 mg/100 g. reported that phytates can reduce It bioavailability of minerals; impaired protein digestibility caused by formation of phytate protein complexes and depressed absorption of nutrients due to damage to the pyloric caeca region of the intestine [63]. Thus, the antinutritional nature of phytic acid lies in its ability to chelate divalent minerals such as iron, calcium, copper and zinc, rendering them biologically unavailable [48]. It also inhibits protein digestion by forming complexes with them [64]. However, the phytate content can further be lowered by processing [65]. The knowledge of the phytate level in foods is necessary because high concentration can cause adverse effects on the digestibility.

Tannins are known to bind irreversibly to proteins of forming insoluble complexes with them and thus rendering them indigestible by intestinal thereby interfering with enzymes their bioavailability [66,67]. According to Jambunathan and Singh, [68], tannins are known to inhibit the activities of digestive enzymes and hence the presence of even a low level of tannin is not desirable from nutritional point of view. Arinathan et al. [69] studied antinutritional factors such as total free phenol, tannin, and hydrogen cynide. Tannin also has the ability to form a complex with vitamin B12 and makes it unavailable [70,63]. They are also known to interact with other antinutrients.

#### 4. CONCLUSION

From the present study, we concluded that flour of ackee (*Blighia sapida*) seeds is a good source of biochemical and nutritional compounds. But the limitation to the full utilization of ackee seeds is the high concentrations of antinutritional factors (oxalate, phytate and tannin) and low alkaloid content which render it useless for human and animal nutrition. However, processing methods, such as boiling and others heat treatment would be better for reducing the levels of these antinutrients present in the flour of ackee raw seeds.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

#### REFERENCES

- 1. Barnett WL. Notes on ackee. Journal of the Jamaica Agricultural Society. 1939;43: 82-85.
- Barceloux DG. Ackee fruit and Jamaican vomiting sickness (*Blighia sapida* Köenig). In Barceloux, D.G. (Ed.) Medical toxicology of natural substances: Foods, fungi, medicinal herbs, toxic plants, and venomous animals. Hoboken (NJ): John Wiley and Sons. 2008;34-38.
- Oyeleke GO, Oyetade OA, Afolabi F, Adegoke BM. Nutrients, antinutrients and physicochemical compositions of *Blighia sapida* pulp and pulp oil (ackee apple). Journal of Applied Chemistry. 2013;4(1): 05-08.
- Benkeblia N, Beaudry RM. Effects of temperature on postharvest respiratory parameters and quality attributes of ackee (*Blighia sapida* Köenig) fruit arils during storage. International Food Research Journal. 2018;25(1):119-126.
- Ekué MRM, Sinsin B, Eyog-Matig O, Finkeldey R. Uses, traditional management, perception of variation and preferences in ackee (*Blighia sapida* K.D. Koenig) fruit traits in Benin: Implications for domestication and conservation. Journal of Ethnobiology and Ethnomedicine. 2010; 6(12):1-14.
- Akintayo ET, Adebayo EA, Arogunde LA. Assessment of dietary exposure to the natural toxin hypoglycin in ackee (Blighia sapida) by Jamaicans. Food Research International. 2002;37:833-838.
- Howélé O, Bobelé N, Théodor D, Séraphi, KC. Nutritional composition studies of sun dried *Blighia sapida* (K. Koenig) aril from Côte d'Ivoire. Journal of Applied Biosciences. 2010;32:1989-1994.
- 8. Veronica MD. Physicochemical and functional properties of different ackee

(blighia sapida) aril flours. Kwame Nkrumah University of Science and Technology. 2014;98.

- AOAC, Official methods of analysis. 15th E dn., Association official analytical chemists. Washington DC. USA; 1990.
- FAO. Carbohydrates in human nutrition. Report of a joint FAO/WHO expert consultation. FAO Food and Nutrition Rome. 1998;66.
- Mathew JT, Ndamitso MM, Otori AA, Shaba EY, Inobeme A, Adamu A. Proximate and mineral compositions of seeds of some conventional and non conventional fruits in Niger State, Nigeria. Acad. Res. Int. 2014;5(2):113-118.
- Dubois M, Mc Cowen LK, Schotch TJ, Rebers PA, Smith F. Colorimetric methods for determination of sugars and related substances. Anales. of Chemistry. 1956;2: 350-356.
- Bernfeld P. Amylase α and β. Methods in enzymology 1.S. P. Colswick and N.O.K., (Eds). Academic Press Inc, New-York. 1955;149-154.
- 14. Pongracz G, Weiser H, Matzinger D. Tocopherols- antioxydant. fat. Science Technology. 1971;97:90-104.
- Barros L, Joao-Ferreira M, Queiros B, Ferreira IC, Baptista P. Food Chem. 2007;413-419.
- Hanson RK, Broom I, Stephenson M. Evaluating community sex offender treatment programs: A 12-year follow-up of 724 offenders. Canadian Journal of Behavioural Science. 2004;36:87-96.
- 17. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. Methods in Enzymology. 1999;299(1):152-178.
- 18. Rodriguez-Amaya DB. A guide to carotenoid analysis in foods. 2001;64.
- 19. Day RA, Underwood AL. Quantitative analysis. 5th Edition, Prentice Hall Publication, Upper Saddle River, 1986;701.
- Bainbridge Z, Tomlins K, Willings K. Westby A. Methods for assessing quality characteristics of non grain starch staple. Part 4 advanced methods. National ressources institute. University of Greenwich, UK. 400–1:43–79. ISBN 0 – 85954 1996
- 21. Meda A, Lamien CE, Romito M, Millogo J. Nacoulma OG. Determination of the total

phenolic, flavonoid and praline contents in burkina fasan honey, as well as their radical scavenging activity. Food Chemistry. 2005;91:571–577.

DOI: 10.1016/j.foodchem.2004.10.006

- 22. Broadhurst RB, Jones WT. Analysis of condensed tannins using acidified vanillin. Journal of the Science of Food and Agriculture. 1978;48(3):788–794.
- Harborne JB. Phytochemical methods: A guide to modern techniques of plant analysis. Chapman and Hall Ltd, London. 1973;279.
- Buratto AP, Carpes ST, Vecchia P, Schroll EM, Appelt P. Determinação da atividade antioxidante e antimicrobiana em castanha-do-pará (Bertholletia excelsa). Revista Brasileira de Pesquisa em Alimentos. 2011;2(1):60-65.
- 25. Lin JY, Tang CY. Determination of total phenolic and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. Food Chemistry. 2007;101(1): 140–147.
- 26. Proestos C, Boziaris IS, Nychas GJE, Komaitis M. Analysis of flavonoids and phenolic acids in Greek aromatic plants: Investigation of their antioxidant capacity and antimicrobial activity. Food Chemistry. 2006;95:664–671.
- Yu F, Cai Y, Kaushik R, Yang X, Chia W. Distinct roles of Galphai and Gbeta13F subunits of the heterotrimeric G protein complex in the mediation of drosophila neuroblast asymmetric divisions. Journal of Cell Biology. 2003;162(4):623-633.
- Maluf MP, Saab IN, Wurtzel ET, Sachs MM. The viviparous12 maize mutant is deficient in abscisic acid, carotenoids, and chlorophyll synthesis. Journal of Experimental Botany. 1997;48:1259–1268.
- 29. Pinzino C, Nanni B, Zandomeneghi M. Aging, free radicals, and antioxidants in wheat seeds. Journal of Agricultural and Food Chemistry. 1999;47: 1333–1339.
- 30. Calucci L, Capocchi A, Galleschi L, Ghiringhelli S, Pinzino C, Saviozzi F, Zandomeneghi M. Antioxidants, free radicals, storage proteins, puroindolines, and proteolytic activities in bread wheat (Triticum aestivum) seeds during accelerated aging. Journal of Agricultural and Food Chemistry. 2004;52:4274-4281.

- 31. Bender DA. Nutritional biochemistry of the vitamins. Cambridge, UK; 2003.
- Fellows PJ. Food processing technology; Principles and Practice, 2<sup>nd</sup> edn. Woodhead Publishing in Food Science and Technology. 2000;238.
- Achinewhu SC. Ascorbic acid content of some Nigeria local fruits and vegetables. Qualities Plantarium Plant Foods for Human Nutrition. 1983;33(4):261-266.
- Adepoju OT. Studies on nutritional characteristics of shea butter (Butyrospernum paradoxum) fruit pulp. Ph.D thesis, department of human nutrition, University of Ibadan, Ibadan, Nigeria; 2004.
- Ogungbenle HN Chemical and fatty acid compositions of date palm fruit (*Phoenix dactylifera* L.) Flour. Bang. J. Sci. Ind. Res. 2011;46(2):255-258.
- Faqir MA, Sardar IB, Ahmad HE, Muhammad IK, Muhammad N, Shahzad H, Muhammad SA. Phytochemical characteristics of date palm (*Phoenix dactylifera*) fruit extracts. Pakistan Journal of Food Science. 2012;22(3):117-127.
- Rehman Z, Salariya AM, Zafar SI. Effect of processing on available carbohydrate content and starch digestibility of kidney beans (*Phaseolus vulgaris* L.). Food Chem. 2012;73:351–355.
- Rodrigues LJO. Pequi (Caryocar brasiliense Camb.): Ciclo vital e agregação de valor pelo processamento mínimo. Lavras, Brasil, Universidade Federal de Lavras, Dissertação de Mestrado; 2005.
- Fennema RO, Tannenbaum SR. Introduction to food chemistry. In: food chemistry, fennema, S.R., M. Karel, G.W. Sanderson, S.R. Tannenbaum, P. Walstra and J.R. Witaker (Eds.). Marcel Dekker Inc., New York. 1996;1-64.
- 40. Aruah BC, Uguru MI, Oyiga BC. Genetic variability and interrelationship among some Nigerian pumpkin accessions (*Curcurbitassp*). Plant Breeding. International Journal. 2012;6:34-41.
- Cust AE, Skilton MR, Van BMME. Total dietary carbohydrate, sugar, starch and fibre intakes in the European Prospective Investigation into cancer and nutrition. Eur. J. Clin. Nutr. 2009;63:37-60.
- Mandalari G, Tomaino A, Arcoraci T, Martorana M, Turco VL, Cacciola F, Rich GT, Bisignano C, Saija A, Dugo P, Cross, KL, Parker ML, Waldron KW, Wickham MSJ. Characterization of polyphenols,

lipids and dietary fibre from almond skins (*Amygdalus communis* L.). J. Food. Comp. Anal. 2010;23(2):166-174.

- 43. Gamal AE, Salah MA, Mutlaq MA. Nutritional quality of biscuit supplemented with wheat bran and date palm fruits (*Phoenix dactylifera* L.). Food Nutritional Sciences. 2012;3:322-328.
- 44. Effiong GS, Ibia IO, Udofia US. Nutritive and energy values of some wild fruit spices in Southeastern Nigeria. Electronic Journal of Environment, Agriculture and food chemistry. 2009;8(10):917-923.
- 45. Ayerza R, Coates W, Lauria M. Chia seed (Salvia hispanica L.) as na omega-3 fatty acid source for broilers: Influence on fatty acid composition, cholesterol and fat content of white and dark meats, growth performance, and sensory characteristics. Poultry Science 2002;816):826-837.
- 46. Enwereuzoh RO, Okafor DC, Uzoukwu AE, Ukanwoke MO, Nwakaudu AA, Uyanwa CN. Flavour extraction from Monodora myristica and Tetrapleura tetraptera and production of flavoured popcorn from the extract. European Journal of Food Science and Technology. 2015;3(2):1.17.
- 47. Lienel HH. Ash analysis, in introduction to chemical analysis of foods, Nielsen, S.S (edn). CBS Publishers, New Delhi. 2002;123-133.
- Food and Agriculture Organization of The United Nations - FAO. Roots, tubers, plantains and bananas in human nutrition. Rome: FAO, (FAO and Food Nutrition series,n. 24). 1990;43.
- 49. Pearson D. The chemical analysis of foods. 7th ed. London: Churchill Livingstone; 1976.
- Dossou VM, Agbenorhevi JK, Combey S. Afi-Koryoe S. Ackee (*Blighia sapida*) fruit arils: Nutritional, phytochemicals and antioxidant properties. 2014;3(6): 534-537.
- 51. Costa GEA, Queiroz–Monici KS, Machado–Reis SMD, Oliveira AC. Chemical composition, dietary fibre and resistant starch content of raw and cooked pea,common bean, chickpea and lentil legumes; 2006.
- 52. Arun AA, Sridhar KR, Raviraja NS, Schmidt E, Jung K. Nutritional and antinutritional components of *Canavalia spp* seeds from the West Coast Sand dunes of India, Plant Food for Human Nutrition. 2004;58:1-13.

Aguemon et al.; AFSJ, 2(2): 1-9, 2018; Article no.AFSJ.40555

- 53. Okaka JC. Cereals and legumes: Storage and processing technology. Data and Microsystems Publishers, Ogui, Enugu, Nigeria. 1997;29-30:12-14.
- 54. Aletor O, Oshodin A, Ipinmoroti KO. Comparative evaluation of the nutritive and psysiochemical characteristics of the leaves and leaf protein concentrates from two edible vegetables. Journal of Food Technology. 2007;5(2):152-156.
- 55. Onyeike EN, Omubo-Dede TT. Effect of heat treatment on the proximate composition, energy values, and levels of some toxicants in African Yam bean (*Sphenostylis stenocarpo*) seed varieties. Plan. Foods Hum. Nutr. 2002;57:223-231.
- 56. Savage GP. Oxalates in human foods. Proceeding of nutrition society. 2002;N2 27:4-24.
- 57. Thompson LU, Yoon JHJ. Starch digestibility as affected by polyphenols and phytic acid. Food Sciences. 1984;49: 1228-1229.
- Nkafamiya II, Modibbo UU, Mariji AJ. Haggai D. Nutrient content of seeds of some wild plants. Afr. J. Biotechnol. 2007;6(14):1665-1669.
- 59. Ojiyako OA, Igwe CU. The nutritive, antinutritive and hepatotoxic properties of Trichosanthes anguina (snake tomato) fruits from Nigeria. Pak. J. Nutr 2008;7: 85-89.
- Bello MO, Farade OS, Adewusi SRA, Olawore NO. Studies of some lesser known Nigerian fruits. African Journal of Biotechnology. 2008;7(1):3972-3979.

- 61. Schjonning P. Christensen BT, Elmhol S. Managing soil quality - Challenges in modern agriculture, CAB International, Wallingford, UK. 2004;69–84
- 62. Aberoumand A. Screening of phytochemical and anti-nutrients compounds of eight food plants sources. World Journal of Science and Technology. 2011;1(4):49–53.
- 63. Francis G, Makkar HPS, Becker R. Antinutritional factors present in plant derived alternative fish feed ingredients and their effects in fish feed ingredients and their effects in fish. Aquaculture L. 2001;99: 197–227.
- 64. Wallingford E. AVI Publishing company, USA. 1985;383-403.
- 65. Eriyamremu GE, Adamso I. Annals of nutrition and metabolism. 1994;38: 174-183.
- 66. Meshansho A, Butler LG, Carbon, DM. Ann. Rev. Nutr. 1987;7:423-430.
- 67. Liener IE. Crit. Rev. Food. Sci. Nutr. 1994;34:31-67.
- Jambunathan R. Singh U. Grain quality of pigeon pea. In: Proceedings of the international workshop on pigeon pea, 15-19 December, 1980, ICRISAT, Hyderabad, Andhra Pradesh, India. 1981;1:351-356.
- 69. Arinathan V, Mohan VR, Maruthupandian A. Tropical Subtropical Agroecosystems 2009;10:273-278.
- Liener IE. Heat labile anti-nutritional factors. In: Advances in legumes sciences (Eds.Summerfield, R.J. and Bunting, A.H). Kew, London, Royal Botanic Gardens. 1980;157–170.

© 2018 Aguemon et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history/24489