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# Effect of Antibiotic and Bio-fungicide for Control of Seed Borne Fungi of Wheat

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## Authors' contributions

This work was carried out in collaboration between all authors. All authors read and approved the final manuscript.

## Article Information

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# ABSTRACT

The present study aimed to evaluate the effect of antibiotic and bio-fungicide for control of seed borne fungi of wheat during November to April. 2015-2016. In this study, two treatments *viz*. Control, Aureofungin (Antibiotic) with 100 ml water, and *Allium sativum* leaf extract with cow urine (Bio-fungicide) for control of seed borne fungi of wheat, In the pot experiment, The result was obtained as a control 63.20%, and disease incidence of 23.30% which were recorded from *Allium sativum* leaf extract with cow urine (Bio-fungicide) in the Treatment 3 whereas control 47.41% and disease incidence 33.33% were recorded from 4 gm. Aureofungin (Antibiotic) with 100 water in the Treatment 2. In the pots, the control was 68.75%, and as disease incidence 20% which were recorded from *Allium sativum* leaf extract with cow urine (Bio-fungicide) in the Treatment 3 whereas control was 52.09% and disease incidence was 30.66% which were recorded from 4 gm. Aureofungin with 100 ml water (Antibiotic) in the T 3 Bio-fungicide was found to be superior in controlling of seed borne fungi of wheat comparing with antibiotic. A bio-fungicide is composed of beneficial microorganisms, such as specialized fungi that attack and control plant pathogens and the diseases.

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## **1. INTRODUCTION**

Antibiotic, the bio-fungicide produces a chemical compound such as an antibiotic or other toxin that kills the target organism. Because of their specificity of action against plant pathogens, relatively low phytotoxicity, absorption through foliage and systemic translocation and activity in low concentration, the use of antibiotic is becoming very popular and very effectively used in managing several plant diseases.

Antibiotic is defined as the inhibition or destruction of the microorganism by substances such as specific or nonspecific metabolites or by the production of anti-biotic that inhibit the growth of another microorganism [1,2,3,4].

Aureofungin is a systemic antifungal antibiotic research product of H.A. (Hindustan Antibiotic) Ltd. It is the only Antifungal Antibiotic in the market. Aureofungin is used for seed treatment as well as for sprays. It is also used for root application. It is either used alone for fungal diseases control or combined with streptocycline when bacterial diseases control is needed.

Seed-borne diseases can seriously affect yield and quality. The most effective means of control is by exclusion and reduction of the inoculums during seed production [5]. Seed-borne diseases have been found to affect the growth and productivity of crop plants [6,7,8]. Seed-borne mycoflora of wheat reported recently included *Alternaria alternata, Drechslera sorokiniana, Fusarium moniliforme, F. avenaceum, F. graminearum, F. nivale, F. culmorum, F. equiseti, F. sporotirchioides, Cladosporium herbarum, Stemphylium botryosum* [9,10,11].

Bio-fungicide means fungicides of biological origin. It may be microorganisms such as bacteria, fungi and animal or plant based product like secondary metabolite. Indian economy is dependent upon agriculture and agriculture has major problems of fungal diseases. Fungi can cause serious damage in agriculture, resulting in critical losses of yield, quality and profit [12].

The present investigation has been undertaken to the study the effect of antibiotic and biofungicide for control of seed borne fungi of wheat with emphasis for further controlling of seed borne fungi using antibiotic and bio-fungicide.

## 2. MATERIALS AND METHODS

The present studies were carried out at Bhargava Agricultural Botany laboratory, Department of Botany, University of Allahabad, Allahabad, Uttar Pradesh, India, during Nov. to April 2015-2016. To evaluate the effect of antibiotic and bio-fungicide in the control of seed borne fungi of wheat.

#### 2.1 Collection of Seed Samples

Seed samples were collected from different seed corporations, companies and farmer's seed lots from two Districts in Allahabad and Varanasi. Then these were properly labeled, kept in polythene bags and stored for further studies in a freezer at 10°C for further studies until ecological testing and other processing [13]. Ten varieties including have been selected, each variety 10 seeds taken in the experiment. Seeds were disinfected with Clorox 1% for 1-2 minutes and then washed three times with distilled water [14].

#### 2.2 Agar Plate Method

Agar plate method [15] as suggested by International Seed Testing Association [16] was used for the isolation of fungi. Seeds were incubated on Agar plates [17]. Potato dextrose agar (PDA) was used for the isolation of mycoflora ten seeds per plate were inoculated and incubated at  $22 \pm 2^{\circ}$ C. After 7 days, incubated seeds were examined under stereo binocular microscope for fungi and then the isolated mycoflora were sub-cultured by single spore technique for macro and microscopic studies.

#### 2.3 Identification of Fungi

Fungal morphology was identified on the basis of colony characteristics and microscopic examinations. Standard books and research papers were consulted during the examination of these fungi [18,19,16]. The fungi were identified with the help of keys, monographs and text provided by several authors [20,21].

#### 2.4 Procedure of Treatments

In the experiment, ten seeds tested with three replications in pot a randomized arrangement and also 100 seeds were tested with three replications in plot. 8 varieties of wheat has been taken as test selected to  $3.06 \times 2.07$  meter plot

Sr. no.	Verity	Market seed	Govt. seed storage	Farmer seed
1	HUW-468	A. niger,	Penicillium spp.,	A. flavus,
		A. flavus	A. flavus	A. niger
2	WH - 147	A. niger,	A. fumigates,	A. niger,
		A. flavus	A. flavus	P. oxalicum
3	Malavashree	A. fumigates,	A. candidus,	A. fumigates,
	(H.I8381)	A. niger	Penicillium spp.	P. oxalicum
4	Gomati (K -9465)	P. griseofulvum,	A. fumigates,	Mucor spp,
		A. fumigatus	P. griseofulvum	Penicillium spp.
5	Prasad (K-8434)	A. niger,	F. oxysporum,	Penicillium spp.,
		F. oxysporum	P. oxalicum	A. flavus
6	Malveey-234	A. flavus,	A. niger,	A. niger,
	-	A. alternate,	A. alternata	C. lunata
		C. lunata		
7	UP-1109	A. niger,	A. niger,	A. fumigates,
		A. ochraceus	A. ochraceus,	A. niger
			A. candidus	
8	R.R21 (Sonalika)	A. fumigates,	A. flavus,	A. flavus,
	. ,	A. niger	A. fumigatus	A. fumigatus
		* (Seth et al. 2015	[22])	

 Table 1. Isolation of fungi from wheat seed, collected from three sites market seed, Govt. seed

 storage and farmer seed in Allahabad

with plant to plant distance 18 cm as a distance between plants and 23 cm between rows [23]. The treatments were respectively applied:-  $T_1$  = Control,  $T_2$  = 3.500 gm Aureofungin with 100 ml water (Antibiotic) at 4 hours for 100 dipping seeds,  $T_3$  = *Allium sativum* leaf extract with cow urine (Bio-fungicide), the wheat seeds were treated from leaf extract of *Allium sativum* with cow urine, dipping seeds in 1:2 ratio preparations [24]. Control percentage and disease incidence of fungal pathogens was calculated by applying these formulas:-

Control % = (Maximum infected seed – Minimum infected seed / Maximum infected seed) × 100

Disease Incidence (D.I.) % = (No. of Infected plants / Total No. of plants) × 100

#### 3. RESULTS AND DISCUSSION

In this experiment, two treatments were applied for controlling the seed borne fungi of wheat in the pots. The result was obtained as a control 63.20%, and disease incidence of 23.30% which were recorded from *Allium sativum* leaf extract with cow urine (Bio-fungicide) in the Treatment 3 whereas control 47.41% and disease incidence 33.33% were recorded from 4 gm. Aureofungin (Antibiotic) with 100 water in the Treatment 2 (Table 2).

The two treatments were also applied for controlling of seed borne fungi of wheat in the

plots. The result was obtained as control 68.75%, and disease incidence 20% which were recorded from *Allium sativum* leaf extract with cow urine (Bio-fungicide) in the Treatment 3 whereas control 52.09% and disease incidence 30.66% were recorded from 4 gm. Aureofungin with 100 ml water (Antibiotic) in the Treatment 3 (Table 3).

It had been observed that, the disease percent had been increased under control treatment. while the treated plants were found to have reduced disease per plant. In a later paper, workers at the same station reported obtaining control of bunt with cycloheximide by means of a 1- minute soaking period, or with dust treatments [25]. Griseofulvin as a antibiotic the causative agent of powdery mildew of wheat, Piricularia oryzae, Ascochyta pisi [26] and silver-leaf disease in plums [27]. Its efficacy has been summarized by [28]. Mycostatin has successfully been used a post-harvest dip-treatment against peach decay [29]. Seeds were treated with the antibiotic aureofungin on Rhizobium strain of groundnut [30]. The treatments of bio-fungicide significantly control percent of seed borne fungi of wheat. They worked on controlling the bacterial leaf blight disease of mango and found that bio-fungicide is an effective control measure and also by [31,32]. The findings of the present study have been supported by [33] as she also found that the bio-fungicide an effective control measure against bacterial leaf blight disease of litchi.

Treatment	Total no. of plants	No. of infected seeds in pots		Mean ±SD	Disease incidence	Control %	
		P <sub>1</sub>	P <sub>2</sub>	P <sub>3</sub>	-	(D.I.%)	
T <sub>1</sub>	10	6	7	6	6.33±0.57	63.33	0.00
T <sub>2</sub>	10	3	2	3	3.33±0.57	33.30	47.41
T <sub>3</sub>	10	2	1	2	2.33±0.57	23.30	63.20

Table 2. Control of seed borne fungi of wheat by different seed treatment in pots

Table 3. Control of	f seed borne fung	gi of wheat b	y different se	ed treatment in	plots

Treatment	Total no. of plants	No. of infected seeds in plots		Mean ± SD	Disease incidence	Control %	
		<b>P</b> <sub>1</sub>	P <sub>2</sub>	P <sub>3</sub>	_	(D.I. %)	
T <sub>1</sub>	100	62	66	64	64.00±2.00	64.00	0.00
T <sub>2</sub>	100	32	30	30	30.66±1.15	30.66	52.09
T <sub>3</sub>	100	22	18	20	20.00±2.00	20.00	68.75

## 4. CONCLUSION

Allium sativum leaf extract with cow urine (Biofungicide) was found to be superior in controlling of seed borne fungi of wheat compared to Aureofungin with 100 ml water (Antibiotic).

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## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# REFERENCES

- Benítez T, Rincón MA, Limón MC, Codón CA. Bio-control mechanisms of *Trichoderma* strains. International Microbiology. 2004;7:249-260.
- 2. Irtwange VS. Application of biological control agents in pre- and postharvest operations. Agricultural Engineering International: The CIGRE Journal. Invited Overview. 2006;3:1-12.
- Haggag WM, Mohamed HAA. Biotechnological aspects of microorganisms used in plant biological control. American-Eurasian Journal of Sustainable Agriculture. 2007;1: 7-12.
- Viterbo A, Inbar J, Hadar Y, Chet I. Plant disease biocontrol and induced resistance via fungal mycoparasites. In: Environ-

mental and Microbial Relationships, 2nd edn. The Mycota IV. (Eds. Kubicek CP, Druzhinina IS). Springer-Verlag Berlin Heidelberg. 2007;127-146.

- Van der Wolf JM, Bimbaum Y, van der Zouwen PS, Groot SPC. Disinfection of vegetable seed by treatment with essential oils, organic acid and plant extracts. Seed Sci. & Technol. 2008;36:76-88.
- Kubiak K, Korbas M. Occurrence of fungal diseases on selected winter wheat cultivars. Postepy w Ochronie Roslin. 1999;39(2):801-804. [In Polish].
- 7. Dawson WAJM, Bateman GL. Fungal communities on roots of wheat and barley and effects of seed treatments containing fluquinconazole applied to control take-all. Plant Pathology. 2001;50:5-82.
- Weber RB, Hrynczuk B, Runowska Hrynczuk, Kita B. Influence of the mode of tillage on diseases of culm base in some winter wheat varieties, oats, and spring wheat. J. Phytopathol. 2001;149:185-188.
- 9. Mirzaj JH, Qureshi MSA. Fungi of Pakistan. Dept. of Plant Pathology. University of Agriculture, Faisalabad, Pakistan. 1978;311.
- Nirenberg H, Schmitz-Elsherif H, Kling CI. Occurrence of *Fusaria* and some "blackening moulds" on durum wheat in (German) Germany. Incidence of *Fusarium* species. Pflanzenkrankheiten und Pflanzenschuntz. 1994;101:449-459.
- Glazek M. Mycoflora of winter wheat seeds harvested from flooded commercial fields in South-Western Poland in 1997. Plant Protection Institute in Poznaniu, Sosnicowice Branch, Gliwicka St. Sosnicowice, Poland. 1997;29:44-153.

Seth et al.; AJEA, 14(1): 1-5, 2016; Article no.AJEA.28988

- 12. Choudhary DK, Verma SK, Patel AK. Dayaram formulation and development of bio-fungicide. International Research Journal of Natural Sciences. 2014;2(2):14-22.
- 13. Fernandez A, Stroshine RL, Tuite J. Mold growth and carbodioxide production during storage of high-moisture corn. Cereal Chem. 1985;62:137-44.
- Mittal RK, Hansen HJ, Thomsen K, Marzalina de M, Khoo de KC, Javanthi de N, Tsna de KFY, Krishnapillav B. Effect of seed treatments and storage temperature on storability of *Syzgium cuminii* seeds. TUFRO Seed Symposium Recalcitrant seeds, Proceedings of the Conference Kaula Lampur Malaysia. 1999;30(1):53-63.
- Agarwal VK. Techniques for the detection of seed borne fungi. Seed Research. 1976;4:24-31.
- Barnet HL, Hunter BB. Illustrated genera of imperfect fungi. The American Psychopathological or Phytopathological Society. 1999;273. U.S.A
- ISTA. Seed health testing methods and the germination test. In. International Rules for Seed Testing. Pub. by Intl. Seed Test. Assoc. Bassersdorf, Switzerland. Spring wheat. J. Phytopathol. 2005;149:185-188.
- Anonymous. International rules for seed testing. Seed Science & Technol. 1993;21: 1-288.
- 19. Rifai M.A. Revision of the genus *Fusarium and Alternaria*. Mycological papers. 1969; 116:40-95.
- 20. Aneja KR. Experiments in microbiology, plant pathology and biotechnology. Fourth Edition, New International (P) Limited Publishers, India. 2004;121-128.
- 21. Barnett HL, Hunter BB. Illustrated genera of imperfect fungi. Burgess Pub. Co., Minneapolis, Minnesota. 1972;241.
- Seth RK, Alam S. Screening of fungi from wheat seeds. International Journal of Agricultural Science and Research (IJASR). 2015;5(5):287-294. ISSN(P): 2250-0057; ISSN(E): 2321-0087.

- 23. Pedro WC, Verkley JM, Groenewald JZ, Samson RA. Fungal biodiversity. Pub. Co. CBSKNAW Fungal. Biodiversity Centre Utrecht, The Netherlands. Phytopathol. 2009;149:185-188.
- Seth RK, Alam S., Shukla DN. Screening of disease resistant varieties against loose smut of wheat J. Nat. Prod. Plant Res. 2014;4(5):49-54.
- 25. Seth RK, Alam S. Indigenous methods: Control of *Septoria* leaf blotch of wheat. International Journal of Agriculture Sciences. 2015;7(13):822-823. ISSN: 0975-3710 & E-ISSN: 0975-9107.
- 26. Henry AW, Millar RL, Peterson EA. Proceedings of the American oriental society meeting at Boston, Mass. American Oriental Society. 1952;115:90-1.
- Wallen VR. Field evaluations and the importance of *Ascochyta* complex on peas. Can. J. Pl. Sci. 1965;45:27-33.
- 28. Bennett M. An approach to the chemotherapy of silver leaf disease (*Stereum purpureum* Fr.) of plum trees. Ann. Appl. Bioi. 1962;50:515-524.
- 29. Rhodes A. Status of griseofulvin in crop protection. Proc. Eastern Sch. Agric. Sci. Univ. Nott. 1962;9:101-124.
- Dimarko GR, Davis BH. Prevention of decay of peaches with postharvest treatment. PI. Dis. Reptr. 1957;41:284-288.
- 31. Mukewar PM, Bhide VP. Effect of seed treatment with fungicides and antibiotic aureofungin on the efficacy of nodulation by *Rhizobium* strain of groundnut. Hindustan Antibiotics Bulletin. 1969;11(3): 172-6.
- 32. Chowdhury MSM. Seed and seedling diseases of some selected fruits of Bangladesh. Ph. D. Thesis. Department of Plant Pathology, Bangladesh Agricultural University, Mymensingh. 2009;97-124.
- 33. Akter H. Management of nursery diseases of Banana and Mango. MS. Thesis. Department of Plant Pathology, Bangladesh Agricultural University, Mymensingh. 2011;60.

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