

European Journal of Medicinal Plants 5(1): 88-94, 2015, Article no.EJMP.2015.007 ISSN: 2231-0894



SCIENCEDOMAIN international www.sciencedomain.org

Petiveria alliacea L (Guinea Hen Weed) and Its Major Metabolite Dibenzyl Trisulfide Demonstrate HIV-1 Reverse Transcriptase Inhibitory Activity

Henry I. C. Lowe^{1,2,3,4*}, Ngeh J. Toyang^{2,3}, Alonso Heredia³, Kenneth N. N. Ayeah², Charah T. Watson¹ and Joseph Bryant³

¹Bio-Tech R&D Institute, Kingston, Jamaica. ²Educational and Scientific Corporation, Wellington, FL, United States. ³Institute of Human Virology, University of Maryland School of Medicine, Baltimore, MD, United States.

⁴College of Health Sciences, University of Technology (UTech), Kingston, Jamaica, West Indies.

Authors' contributions

This work was carried out in collaboration between all authors. Authors HICL, JB and NJT designed the study. Authors NJT and AH carried out the study, wrote the protocol, and wrote the first draft of the manuscript. Authors KNNA and CTW managed the literature searches and with data analysis. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJMP/2015/12064 <u>Editor(s)</u>: (1) Shanfa Lu, Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences & Peking Union Medical College, China. (2) Marcello Iriti, Plant Biology and Pathology Dept. of Agricultural and Environmental Sciences Milan State University, Italy. <u>Reviewers</u>: (1) Anonymous, Cairo University, Egypt. (2) Yared Hailaye, Department of Medicine, School of Medicine, Debre Berhan University, Debre Berhan, Ethiopia. (3) Anonymous, Université Montpellier 1-Mont-pellier 2, Montpellier, France. Peer review History: <u>http://www.sciencedomain.org/review-history.php?iid=648&id=13&aid=6188</u>

> Received 16th June 2014 Accepted 20th August 2014 Published 23rd September 2014

Short Research Article

ABSTRACT

Aim: The human immunodeficiency virus (HIV) remains a major public health concern despite the discovery and development of Highly Active Antiretroviral Therapies (HAART). There is as such a need to continue to search for new and effective therapies for this global pandemic. In an effort to discover new anti HIV agents, the aim of this study was to determine the anti HIV-1 activity of *Petiveria alliacea* and its metabolites.

Methodology: The extracts of *P. alliacea* and dibenzyl trisulfide were screened for anti HIV-1 properties in primary peripheral blood mononuclear cells (PBMCs) infected with the HIV-1JR-CSF strain.

*Corresponding author: Email: henrylowe@cwjamaica.com;

Results: The anticancer metabolite of *P. alliacea* called dibenzyl trisulfide and the crude methanol and ethyl acetate extracts inhibited HIV-1 reverse transcriptase in infected cells, with EC_{50} concentrations of 5.60 µg/ml, 21.6 µg/ml and 68.0 µg/ml, respectively. The reference compound AZT had an EC_{50} value of 0.005 µg/ml. The tested extracts had IC_{50}/EC_{50} selectivity index (SI) values of ≥ 1.47. The results were confirmed in another assay measuring the expression of p24. **Conclusion:** The results of this study indicate that extracts of *P. alliacea* may contain anti HIV-1 metabolites that could provide leads for the discovery of new agents against the HIV virus.

Keywords: Anti HIV-1; Petiveria alliacea; Guinea hen weed; dibenzyl trisulfide; Jamaica; medicinal plant.

1. INTRODUCTION

The Human Immunodeficiency Virus (HIV) is the causative agent Acauired of the Immunodeficiency Syndrome (AIDS) which is considered one of the most important global health problems of the 21st century. Despite the discovery and development of Highly Active Antiretroviral Therapies (HAARTs), the morbidity and mortality due to AIDS remains alarmingly high. Most of the failure to contain the HIV virus has been attributed to several factors including drug resistance [1-3]. Attempts to control the spread of the virus through the development of a vaccine are also yet to yield any viable vaccine candidates [4,5]. The search for new and more active agents against the HIV virus as such remains a priority as existing therapies can become ineffective due to resistance or toxicity [6]. Natural products, especially from plants, have been shown to possess anti HIV activity [7]. The michellamine series of alkaloids, derivatives of betulinic acid and other classes of natural products are amongst some of the molecules isolated from plants with anti HIV activity [8-10]. Jamaica is endowed with several medicinal plant species and is credited for being the source of the Catharanthus roseus plant which yielded vinca alkaloids that were later developed for the treatment of leukemia[11,12].Our research has recently identified a number of Jamaican plants with varying degrees of both anticancer and anti-HIV activity [13,14].

Petiveria alliacea L. is commonly known in Jamaica as the "Guinea Hen Weed", and is claimed to have several medicinal properties. It is used in folk medicine to enhance memory and in the treatment of the common cold, flu, other viral or bacterial infections, inflammation, diabetes, and cancer [15-18]. Studies have confirmed the anticancer potential of guinea Hen Weed *in-vitro* as well as *in-vivo* [19,20]. Dibenzyl trisulfide (DTS) (Fig. 1) was identified as the major metabolite of *P. alliacea* [21], and closer

investigation of its biological properties revealed its anticancer activity, playing a regulatory role in the signal transduction of the mitogen-activated protein kinase (MAPK) pathway [22]. Recent studies have confirmed the involvement of DTS in the MAPK pathway potently inhibiting the ribosomal s6 kinase (RSK), revealing that this might be its mechanisms of action and conferring it with broad cancer therapeutic potential [23]. The first study on the antiviral activity of Guinea Hen Weed was in regard to its activity against the bovine viral diarrhea virus [24]. This paper is focused on the anti HIV-1 activity of Guinea Hen Weed and its anticancer metabolite DTS.

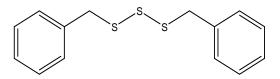


Fig. 1. Chemical structure of dibenzyl trisulfide (DTS)

2. MATERIALS AND METHODS

2.1 Chemicals and Reagents

Major chemicals and reagents where obtained as follows: Dibenzyl trisulfide, 98% purity (International Laboratory, USA); Poly rA:oligodT template: primer (GE Healthcare); DE81 filtermats (Wallac); sodium phosphate buffer or 2X SSC (Life Technologies); MTS CellTiter®96 Reagent (Promega).

2.2 Plant Collection and Extraction

Plant collection: The leaves of *P. alliacea* were collected in Jamaica. A specimen of the plant was identified at the University of the West Indies, Mona Herbarium (deposited under Accession Numbers: IJ 35931). The collected plant material was air dried away from direct sunlight and pulverized.

Plant extraction: *Petiveria alliacea* powder extracted with ethyl acetate and methanol at room temperature for 48 hrs. The filtered solutions were dried in a rotary evaporator. Table 1 presents the details of the extraction and yields.

2.3 Extract Handling

A sample of each extract was dissolved in DMSO and stored at -20°C. The samples were subsequently diluted as necessary prior to use in the anti HIV assay.

2.4 Bioassay

2.4.1 Drugs, cells and virus

Stocks of crude drug made from *P. alliacea* were prepared in DMSO in a stock solution of 10mg/ml. Dilutions from the stock were made in RPMI medium and the final concentration of DMSO in the cultures was always <0.1% to avoid cell toxicity. Peripheral blood mononuclear cells (PBMCs) were separated from buffy coats of healthy donors. Virus used was HIV-1 strains JR-CSF.

2.4.2 Infectivity assays

Three-day phytohemagglutinin (PHA) stimulated PBMCs from at least two normal donors were mixed together. Equivalent number of cells were infected with HIV-1 JRFL using a multiplicity of infection (m.o.i) of 0.001 for 3 h. Infected cells were washed to remove non-adsorbed virus and plated in fresh medium containing IL-2 and various concentrations of drugs. Cells were plated in 96-well plates (2X10⁵ cells/well in 200 µl). Two days after infection, half of the medium was replenished with fresh medium plus drugs. Seven days after infection, supernatants were assayed for virus production by measuring HIV Reverse Transcriptase (RT) activity, and cell viability evaluated.

2.4.3 Reverse transcriptase activity assay

For measuring RT activity, we used apreviously described microtiter plate-based assay [25]. Tritiated thymidine triphosphate (3 H-TTP, 80 Ci/mmol, NEN) was received in 1:1 dH₂O: Ethanol at 1 mCi/mL. Poly rA:oligodT template: primer was prepared as a stock solution by combining 150 µL poly rA (20 mg/mL) with 0.5 mL oligodT (20 units/mL) and 5.35 mL sterile dH₂O, followed by aliquoting (1.0 mL) and

storage at -20°C. The RT reaction buffer was prepared fresh on a daily basis and consisted of 125 μL 1.0 M EGTA, 125 μL dH₂O, 125 μL 20% Triton X100, 50 μL 1.0 M Tris (pH 7.4), 50 μL 1.0 M DTT, and 40 µL 1.0 M MgCl₂. The final reaction mixture was prepared by combining 1 part ³H-TTP, 4 parts dH₂O, 2.5 parts poly rA:oligodT stock and 2.5 parts reaction buffer. Ten microliters of this reaction mixture were placed in a round bottom microtiter plate and 15 µL of virus containing supernatant is added and mixed. The plate was incubated at 37°C for 60 minutes. Following incubation, the reaction volume was spotted onto DE81 filter-mats, washed 5 times for 5 minutes each in a 5% sodium phosphate buffer or 2X SSC, 2 times for 1 minute each in distilled water, 2 times for 1 minute each in 70% ethanol, and then dried. Incorporated radioactivity (counts per minute, CPM) was quantified using standard liquid scintillation techniques. For measuring p24 levels, we used a commercial p24 ELISA assay (Coulter), as previously described [14].

2.4.4 PBMC cytotoxicity assays

At assay termination, plates were stained with the soluble tetrazolium-based dye, MTS Cell Titer®96 Reagent, to determine cell viability. MTS is metabolized by the mitochondria enzymes of metabolically active cells to yield a soluble formazan product, allowing the rapid quantitative analysis of cell viability and compound cytotoxicity. Briefly, 20-25 µL of MTS reagent was added per well, and the microtiter plates were then incubated 4-6 hrs at 37°C, 5% CO₂. Adhesive plate sealers were used in place of the lids, the sealed plate was inverted several times to mix the soluble formazan product and the plate was read spectrophotometrically at 490/650 nm with a Molecular Devices SpectraMax Plus plate reader.

3. RESULTS AND DISCUSSION

The yields of the different extracts of *P. alliacea* are presented in Table 1. The methanol extract had the highest yield (4.646%) and the ethyl acetate extract had a yield of 0.370% which was only about 8% of the methanol extract yield.

The results of the *P. alliacea* reverse transcriptase (RT) activity against the HIV-1 1JR-CSF strain are presented in Table 2 and Fig. 2. The cytotoxic effects of these extracts on PBMCs were measured simultaneously in the assay to ascertain that RT effect was not influenced by

cytotoxicity. The methanolic extract demonstrated higher activity compared to the ethyl acetate extract in the RT assay, with an EC_{50} of 21.60 µg/ml and an IC_{50} >100 µg/ml. The SI for the methanol extract was >4.62, compared to an SI of only >1.47for the ethyl acetate extract. Similar results were obtained by measuring p24 levels in the culture supernatants (data not shown).

In contrast, DTS was active against HIV-1 JRC-SF but had higher cytotoxicity compared to the crude extracts. Being a pure molecule, DTS is as such unlikely to be regarded as a possible anti HIV-1 agent given its poor selectivity. The implication is that there is possibly a different molecule(s) responsible for the anti HIV-1 activity of the crude extracts of *P. alliacea*. The fact that the methanolic extract exhibits antiviral activity in the RT assay suggests that the anti HIV-1 molecule(s) might be of mid to high polarity given that DTS is highly nonpolar.

Table 1. Extraction of *P. alliacea* using different solvents

Solvent	Volume	Amount (g)	Yield (g)	% Yield
Methanol	500 mL	25	1.162	4.646
Ethyl	500 mL	25	0.093	0.370
acetate				

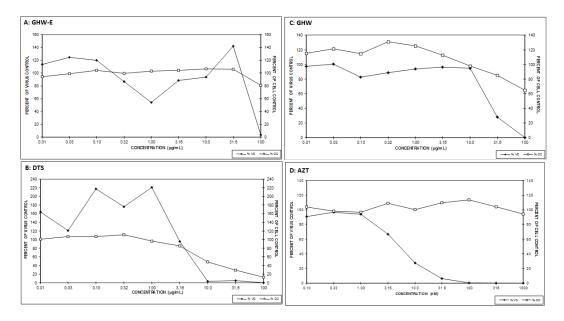


Fig. 2. Inhibition of HIV-1 JR-CSF replication in PBMC by *P. alliacea* extracts and its major metabolite DTS. AZT (D) was used as a positive control. HIV-1 replication inhibition was simultaneously determined with the cytotoxicity of the test samples on PBMC cells. For the HIV-1 replication inhibition assay, three-day phytohemagglutinin (PHA) stimulated PBMCs from at least two normal donors were mixed together and used. Seven days after infection, supernatants were assayed for virus production by measuring HIV Reverse Transcriptase (RT) activity. To determine cytotoxicity, plates were stained with the soluble tetrazolium-based dye, MTS CellTiter®96 Reagent, to determine cell viability

Table 2. Anti HIV-1 activity of extracts of P. alliacea L. and dibenzyl trisulfide (DTS)

Extract/compound	Activity against		Selectivity index (SI)	
	EC ₅₀ (μg/ml)	IC ₅₀ (μg/ml)		
GHW	21.60	>100	>4.62	
GHW-E	68.00	>100	>1.47	
DTS	5.60	9.53	1.70	
AZT	0.005	>1	>230	

 $SI = IC_{50}/EC_{50}$. EC_{50} and IC_{50} values were determined by variable slope non-linear regression analysis of the plotted data using GraphPad Prism software. P. alliacea extracts: ethyl acetate (GHW-E) and methanol (GHW)

While the possible mechanism of action of P. alliacea extracts against the HIV-1 virus remains to be discovered, data from previous studies indicate that P. alliacea extracts do decrease the expression of cyclophylin A in a breast carcinoma cell line (4T1) [19]. This is very interesting because human cyclophilins A and B are host cell proteins that bind specifically to the HIV-1 Gag polyprotein p55^{gag} in vitro and drugin virion-associated induced reductions cyclophilin A levels have been shown to be accompanied by reductions in HIV-1 virion infectivity, indicating that the association is functionally relevant for HIV-1 infectivity [26,27]. Based on the findings that cyclophilins are important for viral capsid assembly, cyclophilins are as such considered drugable targets in antiviral therapy [28,29]. Other studies that lend support to the therapeutic potential of P. alliacea against HIV-1/AIDS is the fact that this plant is credited for having immunomodulatory properties [21,30]. The human immune system is used to fight off infections and as such it is one of the first targets attacked by the HIV-1 virus guaranteeing its survival in the human system [31,32]. To combat the HIV infection and slow the progression to full blown AIDS, immunotherapy is often recommended as a therapeutic strategy [32-34]. Stimulation of natural immune response to infections are considered necessary for the development of an HIV vaccine [35]. An antioxidant metabolite was also isolated and characterized from P. alliacea yielding further evidence of the immunomodulatory properties of this plant [36].

4. CONCLUSION

In conclusion, the plethora of evidence on the medicinal properties of P. alliacea keeps growing, making this plant one of the most popular medicinal plants in Jamaica, and the Caribbean Islands in general. The anti HIV-1 activity of P. alliacea is reported herein for the first time. Given that this plant is already being marketed in Jamaica in capsules and as a tea, it is possible that HIV infected persons taking the supplements for other reasons might be benefiting from the anti HIV effects of this plant without knowing. The existing information on its possible mechanism of action, combined with its reported effects on the immune system, makes this plant a good candidate for the potential development of a new therapy to be used alone or in combination with other agents against HIV-1. The activity of P. alliacea against reference HIV-1 strain JRCSF, which uses CCR5 as a coreceptor, is important in that CCR5 strains are the predominantly transmitted and generally persist during the course of infection [37-40]. In summary, our data suggest that *P. alliacea* may have therapeutic potential against HIV-1 and further studies are required to demonstrate the extent of its activity.

CONSENT

Not applicable.

ETHICAL CLEARANCE

Not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Organization WH. The HIV drug resistance report-2012: World Health Organization; 2012.
- Fofana D, Soulié C, Baldé A, et al. High level of HIV-1 resistance in patients failing long-term first-line antiretroviral therapy in Mali. Journal of Antimicrobial Chemotherapy. 2014:dku153.
- Wainberg MA, Zaharatos GJ, Brenner BG. Development of antiretroviral drug resistance. New England Journal of Medicine. 2011;365:637-46.
- Sekaly RP. The failed HIV Merck vaccine study: A step back or a launching point for future vaccine development? The Journal of Experimental Medicine. 2008;205:7-12.
- Verkoczy L, Kelsoe G, Haynes BF. HIV-1 Envelope gp41 Broadly Neutralizing Antibodies: Hurdles for Vaccine Development. PLoS Pathogens. 2014;10:e1004073.
- Mendizabal-Burastero R, Girón-Callejas AC, Rodas-Cruz JA, et al. [Patterns of HIV-1 resistance to antiretroviral drugs in patients with virologic failure: Roosevelt Hospital, Guatemala 2008-2012]. Revista Chilena De Infectious Diseases: Official Publication of the Infectious Diseases Society of Chile. 2013;30:630-7.
- Cragg G, Newman D. Plants as a source of anti-cancer and anti-HIV agents. Annals of Applied Biology. 2003;143:127-33.

- Boyd MR, Hallock YF, Cardellina JH, et al. Anti-HIV michellamines from Ancistrocladus korupensis. Journal of Medicinal Chemistry. 1994;37:1740-5.
- Fujioka T, Kashiwada Y, Kilkuskie RE, et al. Anti-AIDS agents, 11. Betulinic acid and platanic acid as anti-HIV principles from Syzigium claviflorum, and the anti-HIV activity of structurally related triterpenoids. Journal of Natural Products. 1994;57:243-7.
- Asada Y, Sukemori A, Watanabe T, et al. Stelleralides A–C, Novel Potent Anti-HIV Daphnane-Type Diterpenoids from Stellera chamaejasm e L. Organic Letters. 2011;13:2904-7.
- 11. Noble RL. The discovery of the vinca alkaloids-chemotherapeutic agents against cancer. Biochemistry and Cell Biology. 1990;68:1344-51.
- 12. Wright JR. Almost famous: E. Clark Noble, the common thread in the discovery of insulin and vinblastine. Canadian Medical Association Journal. 2002;167:1391-6.
- 13. Lowe H, Toyang N, Bryant J. *In vitro* and *In vivo* anti-cancer effects of tillandsia recurvata (Ball Moss) from Jamaica. West Indian Medical Journal. 2013;62.
- Lowe HI, Toyang NJ, Heredia A, et al. Anti HIV-1 Activity of the Crude Extracts of *Guaiacum officinale* L.(*Zygophyllaceae*). European Journal of Medicinal Plants. 2014;4.
- Williams L, Rosner H, Levy H, Barton E. A critical review of the therapeutic potential of dibenzyl trisulphide isolated from *Petiveria alliacea* L (Guinea hen weed, anamu). West Indian Medical Journal. 2007;56:17-21.
- Kim S, Kubec R, Musah RA. Antibacterial and antifungal activity of sulfur-containing compounds from *Petiveria alliacea* L. Journal of Ethnopharmacology. 2006;104:188-92.
- 17. Lopes-Martins R, Pegoraro D, Woisky R et al. The anti-Inflammatory and analgesic effects of a crude extract of *Petiveria alliacea* L. (*Phytolaccaceae*). Phytomedicine. 2002;9:245-8.
- Gomes PB, Oliveira MMdS, Nogueira CRA, et al. Study of antinociceptive effect of isolated fractions from *Petiveria alliacea* L.(tipi) in mice. Biol Pharm Bull. 2005;28:42-6.
- 19. Hernández JF, Urueña CP, Cifuentes MC, et al. A *Petiveria alliacea* standardized fraction induces breast adenocarcinoma

cell death by modulating glycolytic metabolism. Journal of Ethnopharmacology. 2014;153:641-9.

- 20. Cifuentes MC, Castañeda DM, Urueña CP, Fiorentino S. A fraction from *Petiveria alliacea* induces apoptosis via a mitochondria-dependent pathway and regulates HSP70 expression. Universitas Scientiarum. 2011;14:125-34.
- 21. Williams LA, The T, Gardner M, et al. Immunomodulatory activities of *Petiveria alliacea* L. Phytotherapy Research. 1997;11:251-3.
- Rösner H, Williams L, Jung A, Kraus W. Disassembly of microtubules and inhibition of neurite outgrowth, neuroblastoma cell proliferation, and MAP kinase tyrosine dephosphorylation by dibenzyl trisulphide. Biochimica et Biophysica Acta (BBA)-Molecular Cell Research. 2001;1540:166-77.
- 23. Lowe HI, Facey CO, Toyang NJ, Bryant JL. Specific RSK Kinase Inhibition by Dibenzyl Trisulfide and Implication for Therapeutic Treatment of Cancer. Anticancer Research. 2014;34:1637-41.
- 24. Ruffa M, Perusina M, Alfonso V, et al. Antiviral activity of *Petiveria alliacea* against the bovine viral diarrhea virus. Chemotherapy. 2002;48:144-7.
- Buckheit Jr RW, Swanstrom R. Characterization of an HIV-1 isolate displaying an apparent absence of virionassociated reverse transcriptase activity. AIDS Research and Human Retroviruses. 1991;7:295-302.
- 26. Thali M, Bukovsky A, Kondo E, et al. Functional association of cyclophilin A with HIV-1 virions. Nature. 1994;372:363-5.
- 27. Franke EK, Yuan HEH, Luban J. Specific incorporation of cyclophilin A into HIV-1 virions. Nature. 1994;372:359-62.
- Flisiak R, Horban A, Gallay P, et al. The cyclophilin inhibitor Debio-025 shows potent anti-hepatitis C effect in patients coinfected with hepatitis C and human immunodeficiency virus. Hepatology. 2008;47:817-26.
- 29. Moore JP, Stevenson M. New targets for inhibitors of HIV-1 replication. Nature Reviews Molecular Cell Biology. 2000;1:40-9.
- 30. Santander SP, Hernández JF, Cifuentes BC, et al. Immunomodulatory effects of aqueous and organic fractions from *Petiveria alliacea* on human dendritic cells.

The American Journal of Chinese Medicine. 2012;40:833-44.

- Abbas AK, Lichtman AH, Pillai S. Basic immunology: Functions and disorders of the immune system: Elsevier Health Sciences; 2012.
- 32. Letvin NL, Walker BD. Immunopathogenesis and immunotherapy in AIDS virus infections. Nature Medicine. 2003;9.
- Phimister EG, Weiss RA. Immunotherapy for HIV Infection. New England Journal of Medicine. 2014;370:379-80.
- Weiss RA. Immunotherapy for HIV Infection. The New England Journal of Medicine. 2014;370:379-80.
- 35. Johnston MI, Fauci AS. An HIV vaccine challenges and prospects. New England Journal of Medicine. 2008;359:888-90.
- Okada Y, Tanaka K, Sato E, Okajima H. Antioxidant activity of the new thiosulfinate

derivative, S-benzyl phenylmethanethiosulfinate, from *Petiveria alliacea* L. Organic & Biomolecular Chemistry. 2008;6:1097-102.

- Michael NL, Chang G, Louie LG, et al. The role of viral phenotype and CCR-5 gene defects in HIV-1 transmission and disease progression. Nat Med. 1997;3:338-40.
- Michael NL, Louie LG, Rohrbaugh AL, et al. The role of CCR5 and CCR2 polymorphisms in HIV-1 transmission and disease progression. Nat Med. 1997;3:1160-2.
- Wilen CB, Tilton JC, Doms RW. HIV: Cell binding and entry. Cold Spring Harbor Perspectives in Medicine. 2012;2.
- 40. Shaw GM, Hunter E. HIV transmission. Cold Spring Harbor Perspectives in Medicine. 2012;2.

© 2015 Lowe et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history.php?iid=648&id=13&aid=6188