



Evaluation of Phytochemicals and Biological Activity of *Silybum marianum* L.

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Author's contribution

The sole author designed, analyzed and interpreted and prepared the manuscript.

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ABSTRACT

Aims: The goal of this work is the evaluation of phytochemicals and biological activity of *Silybum marianum*.

Place and Duration of Study: *S. marianum* L. and seeds of *Chenopodium murale* L. were collected from different sites from Al Anbar, Iraq during the period of May 2017.

Methodology: Total phenolics, tannins, alkaloids, flavonoids and saponins, were determined. The antioxidant activity was measured based on the reduction of DPPH (1,1-diphenyl-2-picrylhydrazyl). Antibacterial activity of the extracts was determined against ten microorganisms using the disc diffusion method as well as the allelopathic potential of the *S. marianum* extract on the germination of *C. murale* was studied.

Results: The IC₅₀ values of Methanol extract of *S. marianum* extracts had the highest scavenging activity (1.99 mg.ml⁻¹). The radical scavenging activity of the other extracts and standard decreased in the following order: catechol, ethyl acetate, hexane and petroleum ether. In the present study the *S. marianum* extracts exhibited different inhibitory activities against the tested microorganisms with different degrees, methanol and ethyl acetate extracts of *S. marianum* showed the broad spectrum against the tested microorganisms. The pathogen *Escherichia coli* was the most sensitive bacteria (25 mm), while *Aspergillus fumigatus* and *A. niger* were the most sensitive fungi (25 and 27 mm) in case of methanol extract. The phytotoxicity of *S. marianum* extracts was increased significantly with

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the increasing extract concentrations. At 25 g.l⁻¹ and 20 g.l⁻¹ the germination of *C. murale* was reached maximum inhibition (92.45% and 76.84, respectively). In case of *C. murale* radicle growth, the higher concentration (25 g.l⁻¹) was strongly inhibited (97.23%) the *C. murale*. Similarity, the plant extract significantly reduced the plumule length of *C. murale*. The highest concentrations (25 g.l⁻¹) of extract showed strong inhibition (87.29%) of plumule growth.

Conclusion: *S. marianum* extracts can be used as natural antioxidant, antimicrobial agents pharmaceutical and bio-control of weeds.

Keywords: *Silybum marianum*; Asteraceae; antioxidant; antimicrobial; phytotoxicity; Iraq.

1. INTRODUCTION

The utilization of plant and its products has a long history that began with folk medicine and through the years has been incorporated into traditional and allopathic medicine. Medicinal plants are of play vital importance to the health of individuals and communities. Herbal medicines derived from plant extracts are being increasingly utilized in traditional treatments for many human diseases for thousands of years and in many regions of the world [1]. Plants provide abundant resources of antimicrobial compounds and have been used for centuries to inhibit microbial growth [2].

The medicinal value of many plants species reported having pharmacological properties due to the presence of various kinds of phytochemicals including alkaloids, flavonoids, glycosides, saponins, steroids, terpenes and other phenolic compounds which are therefore, should be utilized to against the disease-causing pathogens [3,4]. The screening of the nutritional composition as the relevance of the presence of phytochemical and antioxidative potentials in a wild plant. Antioxidants or inhibitors of oxidation are compounds which retard or inhibit the oxidation and in general prolong the life of the oxidizable matter [5,6].

Silybum marianum (milk thistle, family Asteraceae) is an annual or biennial herb. Its stem is 20 to 150 cm in height while its leaves ranged from 25 to 50 cm lengthy and 12 to 25 cm wide. The fruit is hard-skinned achene with brown spots and is 15 to 20 mm lengthy [7]. *S. marianum* is native to southern Europe, mainly the Mediterranean region, indigenous to Asia, Southern Europe, North America, and Russian Federation. It is naturalized in South and North America, Australia, China, Central Europe [8]. *S. marianum* is a wild growing annually herb that grows in many parts of the world including the

north part of Iraq and some area north Baghdad city [9].

The plant is known for its medicinal properties having essential biochemical constituents including many flavonolignans collectively known as silymarin. Silymarin has antioxidant properties and utilized as part of hepatic disorders, including hepatotoxicity secondary to acute and chronic viral hepatitis and mushroom poisoning [10]. The present study was conducted to evaluate phytochemicals and biological activity of *S. marianum*.

2. MATERIALS AND METHODS

2.1 Plant Material and Preparation of the Extract

Silybum marianum L. was collected from different sites from Al Anbar, Iraq during the period of May 2017. The identification of species was done according to Boulos [11]. It was dried at room temperature and grinded into a powder using a blender. Ten gram of dried plant powder was extracted using different solvents (Methanol, hexane, petroleum ether and ethyl acetate) by soaking overnight with periodical shacking. The solution was filtered and evaporated to dryness. The dried residue was dissolved in dimethyl sulfoxide (DMSO) and preserved at -20°C for future use [12].

2.2 Phytochemical Analysis

S. marianum was collected and prepared as previously mentioned. Total phenolics, flavonoids and alkaloids were estimated using spectrophotometric techniques adapted by Harborne [13], Sadasivam and Manickam [14] and Boham and Kocipai-Abyazan [15], respectively. Tannins were determined according to Van-Buren and Robinson [16], while saponin amount was estimated by the method adopted by Obadoni and Ochuko [17].

2.3 Biological Activity

2.3.1 Evaluation of DPPH free radical scavenging activity

Antioxidant activity was determined by using a stable free radical (1,1-diphenyl-2-picrylhydrazyl) DPPH [18]. Two ml of 0.15 mM DPPH was added to 2 ml of plant extracts in different concentrations (4000, 2000, 1000, 500 and 250 ppm). A control was prepared by adding 2 ml of DPPH to 2 ml solvent. The mixture was kept in dark at 37°C for 30 min. The absorbance was recorded at 517 nm and the IC₅₀ was calculated graphically. The antioxidant activity was expressed as:

$$\% \text{ Radical scavenging activity} = [1 - (A_{\text{sample}}/A_{\text{control}})] \times 100$$

2.3.2 Antimicrobial bioassay

2.3.2.1 Antibacterial activity

Antibacterial activity of the extracts was determined against seven bacterial strains, three gram-positive i.e., *Streptococcus pyogenes*, *Staphylococcus aureus* and *Bacillus subtilis* and four gram-negative i.e., *Proteus vulgaris*, *Klebsiella pneumoniae*, *Shigella* and *Escherichia coli* using the disc diffusion method. A sterile paper disc (5 mm in diameter) was soaked in the crude extract of the studied plant and then placed over the surface of the inoculated nutrient agar in antibacterial assay [19]. All Petri dishes were incubated at 37°C for 24 hrs. After incubation, the diameter of inhibition zone (cm) was measured for recording the clear zone and compared with the DMSO as a control. Experiments were performed in triplicate and mean inhibitory zone was calculated. The standard antibiotic of ampicillin, clotrimazole and penicillin were used for comparison with the tested plant extracts.

2.3.2.2 Antifungal activity

Antifungal activity against three fungal strains (*Aspergillus fumigatus*, *Aspergillus niger* and *Candida albicans*) was determined by the disc diffusion [19]. Filter paper discs (5 mm in diameter) are prepared before use and sterilized in an autoclave for 20-30 min. A sterile paper disc is wetted in the solution of crude extract (100 µl) and then placed over the surface of the inoculated PDA in the antifungal assay described by Culture plates were incubated at 28°C for

72 hrs. and zones of inhibition was recorded around the paper disc.

2.3.3 Allelopathic bioassay

The seeds of *Chenopodium murale* were collected from the cultivated land from Al Anbar, Iraq. Seeds were sterilized by 0.3% sodium hypochlorite for 3 minutes, washed several times with distilled water, dried at room temperature for 7 days and reserved in a paper bag until further use [20]. For bioassay assessments, methanol extracts were set up at distinct concentrations (2.5, 5, 10, 20% w/v). The solutions had been filtered via double layers of muslin cloth followed by a Whatman No. 1 filter paper. The pH of the mixtures was adjusted to 7 with 1M HCl, and then mixtures have been kept in a refrigerator at 4°C till additionally utilize [21].

For germination experiment, two layers of filter paper (Whatman No. 1) were placed in 90 mm diameter sterilized Petri dishes. In each dish, 20 seeds were settled and 10 ml of each plant extract (2.5, 5, 10, 20% w/v) was added. The control treatment was designed with distilled water. Germinated seeds were counted daily starting from the first day of treatment. The design of the experiment was a randomized complete block with three replicates. The experiment repeated three times and the inhibition percentage was calculated.

The seeds of *C. murale* were germinated in the dark at room temperature for 48 hrs. Twenty germinated seeds were placed in Petri dishes lined with two layers of filter paper (Whatman No. 1) and 10 ml of different extracts (2.5, 5, 10, 20% w/v) were added. Moreover, a control treatment was designed with distilled water. The design of the experiment was a randomized complete block with three replicates. The experiment repeated twice, the radicle and plumule lengths of seedlings were measured on the tenth day and growth inhibition for radicle and plumule lengths were calculated.

3. RESULTS AND DISCUSSION

3.1 Phytochemical Constituents

Phytochemicals are playing a vital role for the treatment of different types of diseases and still are used in, both traditional and modern system of medication. The phytochemical analysis of the aerial parts of *S. marianum* indicated that the plant is rich in secondary compounds. The

results indicated that the *S. marianum* exhibited the highest content of tannins and phenolics (11.42 ± 0.65 and 16.3 ± 1.07 mg.g^{-1} , respectively), followed by saponins (9.54 ± 0.3 mg.g^{-1}), flavonoids (9.12 ± 0.52 mg.g^{-1}) and then alkaloids (7.33 ± 0.5 mg.g^{-1}). This result is supported by the study of Shah et al. [22] and Salem et al. [23]. In addition, this results relatively comparable to those reported in *Senecio glaucus* and *Urospermum picroides* as described by El-Amier et al. [24,25].

3.2 Antioxidant Activity

The assessment of the antioxidant activity of the diverse plant extracts is presented in Fig. 1. By increasing the plant extract concentration there was a corresponding continuous increase in scavenging activity. In case of methanol, ethyl acetate, petroleum ether and hexane extracts the increase was up to $4000 \mu\text{g.ml}^{-1}$ where the scavenging activity was 53.52%, 50.48%, 37.53% and 36.39, respectively. The IC_{50} values of *S. marianum* extracts were presented in Fig. 2. Methanol extract had the highest scavenging activity (1.99 mg.ml^{-1}). The radical scavenging activity of the other extracts and standard decreased in the following order: catechol, ethyl acetate, hexane and petroleum ether. These results suggest that methanol extract of *S. marianum* has an obvious effect on scavenging of DPPH radical. Similar results were reported by Salem et al. [23] and El-Amier & Abdullah [26], while investigating the DPPH radical scavenging

activity of *S. marianum* and *Senecio glaucus*, respectively.

3.3 Antimicrobial Activity Assessment

Microbial disease and contamination have become one of the main problems of public health in the world, affecting all countries. It can be connected to the process of natural selection in bacterial development or the natural consequence of the adaptation of pathogen to exposure to antibiotics in the course of the indiscriminate use of antibiotics in humans and animals [27]. The antibacterial activity of *S. marianum* was assayed in vitro by agar well diffusion method against seven different bacterial strains. The part used for the study was shoot system and four extracts (methanol, petroleum ether, hexane and ethyl acetate) were evaluated for antimicrobial activity as shown in Table 2 and Fig. 3. *S. marianum* extracts exhibited different inhibitory activities against the tested bacterial and fungal strains with different degrees as demonstrated by measuring the diameters of inhibition zones developed by the extracts. Table 1 showed that methanol and ethyl acetate extracts of *S. marianum* showed the broad spectrum against the tested bacteria Fig. 4. Whereas, hexane extracts expressed an activity against *Shigella* spp and *P. vulgaris*, only. Petroleum ether extract showed the inhibitory activities against *E. coli*, *Klebsiella pneumoniae* and *Candida albicans*.

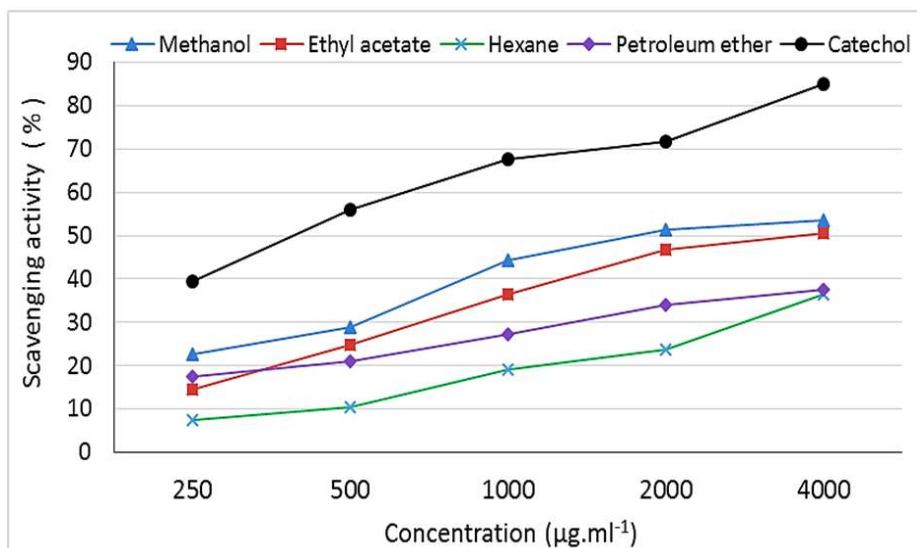
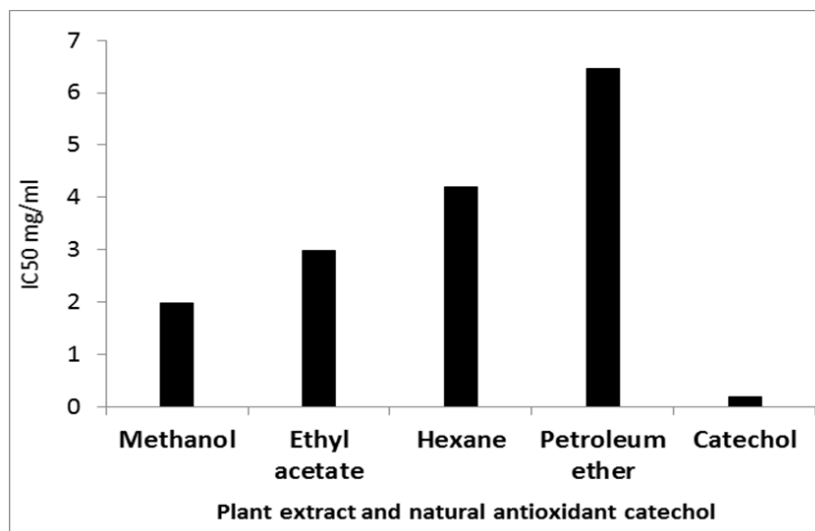


Fig. 1. % of scavenging activity of *Silybum marianum* extracts and natural antioxidant catechol

Table 1. The concentration of the active constituents in mg/g dry weight for the *S. marianum*

Plant species	mg/g Dry weight				
	Tannins	Saponins	Alkaloids	Flavonoids	Phenolics
<i>S. marianum</i>	11.42±0.65	9.54±0.3	7.33±0.5	9.12±0.52	16.3±1.07

**Fig. 2. IC₅₀ values (mg.ml⁻¹) of *Silybum marianum* extracts and natural antioxidant catechol (standard)**

S. marianum produced inhibition zones less than that of the standard antibiotic ampicillin and clotrimazole against tested organisms. The pathogen *E. coli* was the most sensitive bacteria (25 mm), while *A. fumigatus* and *A. niger* were the most sensitive fungi (25 and 27 mm) in case of methanol extract. The present results agree with those of Izzo et al. [28] and Dayanne et al. [27] on same species. Ramdani et al. [29] found similar results that the methanol extract of *Urospermum dalechampii* from Algeria was active against *E. coli*, *K. pneumoniae*, *P. aeruginosa*, *B. subtilis* and *S. aureus*. El-Amier et al. [24] also reported that *Sencio glaucus* (Asteraceae) extract showed an inhibition zone against *E. carotovora*, *S. biogenesis* and *B. subtilis* but not against *S. aureus*, *E. coli* and *P. aeruginosa*. The main constituents of phytochemical analysis of *S. marianum* are silibinin, isosilibinin, silicristin, and silidianin (Sonnenbichler et al. 1999). Silymarin has been found very active against most microorganisms [30,27].

3.4 Allelopathic Activity

The allelopathic potential of the *S. marianum* extract on the germination of *C. murale* 4 days

after treatment is presented in Fig. 5. The phytotoxicity of extracts was increased significantly with the increasing extract concentrations. At 25 g.l⁻¹ and 20 g.l⁻¹ the germination of *C. murale* was reached maximum inhibition (92.45% and 76.84, respectively). However, at the lowest concentration (5 g.l⁻¹), the germination of *C. murale* was reduced by 14.26%. This agrees with the previous results of other investigators [31,32].

On the other hand, The allelopathic effect of the different concentration from extract of *S. marianum* on *C. murale* radicle growth after 10 DAT revealed that the higher concentration (25 g.l⁻¹) was strongly inhibited (97.23%) the *C. murale*, while the opposite response (26.17%) was observed at the lower concentration (5 g.l⁻¹) (Fig. 5). Similarity, the plant extract significantly reduced the plumule length of *C. murale* (Fig. 5). The highest concentrations (25 g.l⁻¹) of extract showed strong inhibition (87.29%) of plumule growth. The results of the prevailing studies agree with most of the previous results obtained by other researchers, which emphasized that extracts of many plant species inhibited germination of many other weed seeds [33,34,31].

Table 2. The inhibitory activity of the plant extract against the tested organisms as demonstrated by diameters of the inhibition zone (mm)*

Test microorganisms	Plant extracts				Standard antibiotic	
	Methyl alcohol	Ethyl acetate	Hexane	Petroleum ether	Ampicillin	Clotrimazole
Gram-positive bacteria						
<i>B. subtilis</i>	-	22	-	-	-	-
<i>S. aureus</i>	21	19	-	-	32	-
<i>S. pyogenes</i>	-	-	-	-	25	-
Gram-negative bacteria						
<i>E. coli</i>	25	21	-	9	36	-
<i>K. pneumonia</i>	20	-	-	10	29	-
<i>Shigella spp</i>	10	-	4	-	-	-
<i>P. vulgaris</i>	-	13	10	-	16	-
Fungi						
<i>C. albicans</i>	10	11	-	14	-	15
<i>A. niger</i>	25	18	-	-	-	33
<i>A. fumigatus</i>	27	15	-	-	-	31

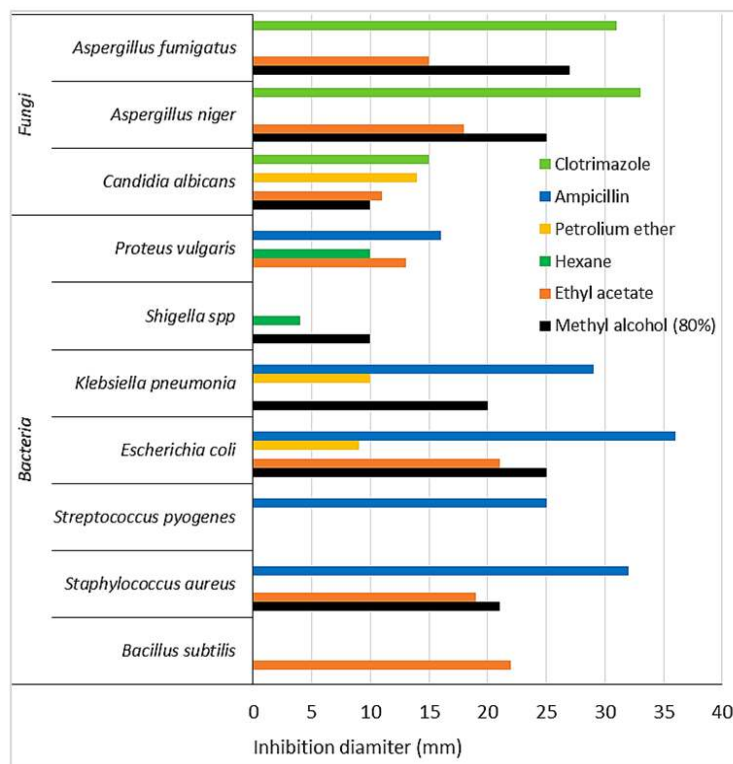


Fig. 3. Antimicrobial activity of different extract of *S. marianum* and standard antibiotic

It was noticeable that the degree of inhibition percentage of the plant extracts increased with the increase in its concentration. The present results showed the potent allelopathic effect of *S. marianum* on the nuisance weed *C. murale*, which could be ascribed to the high content of phenolics, tannins and alkaloids. The reduction in

the germination of weeds could be attributed to the action of allelochemicals in the plant. Allelochemicals pose great effects on the membrane permeability, enzyme activities, cell division and ultrastructure, ion uptake and as a consequence germination, plant growth and development are modified [35,36].

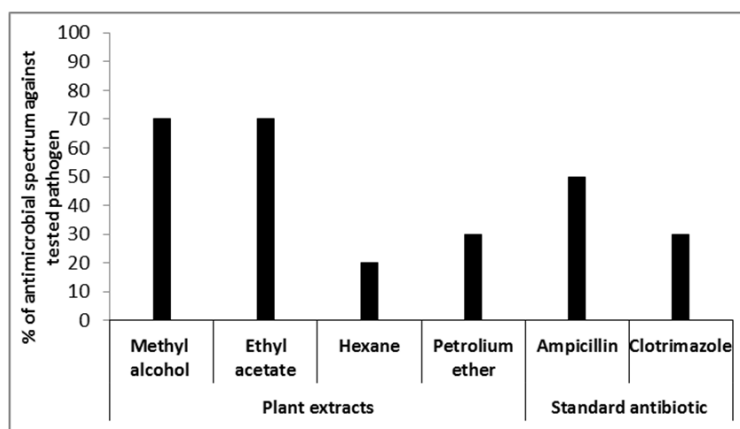


Fig. 4. % of antimicrobial spectrum of *S. marianum* extracts and standard antibiotic

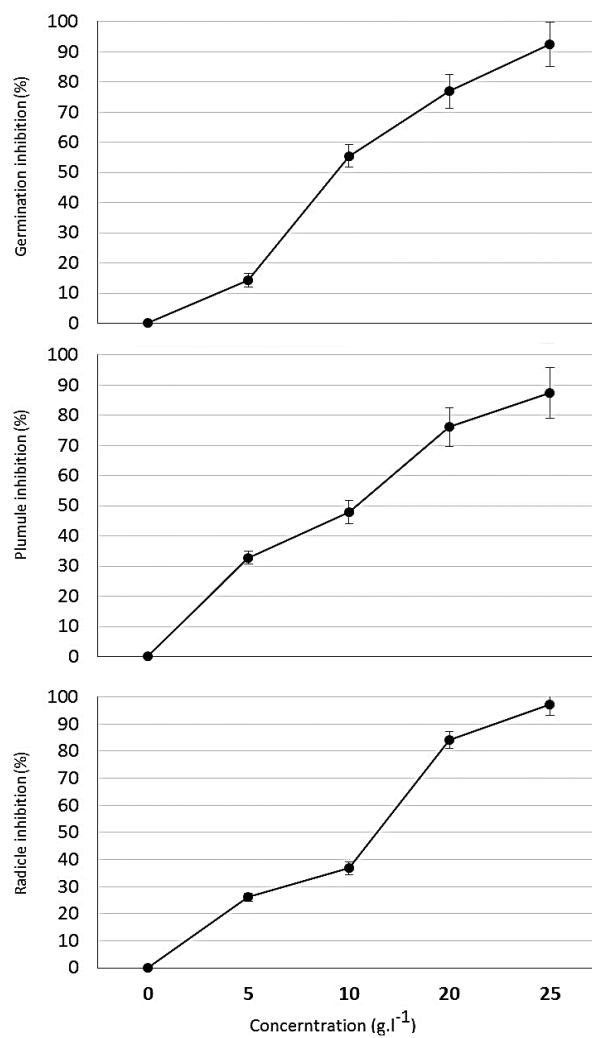


Fig. 5. Allelopathic effect of different methanol extracts from *S. marianum* aerial parts on the germination and seedling growth of *C. murale* after ten days of treatment

4. CONCLUSION

In the present study, the *S. marianum* extracts exhibited different inhibitory activities against the tested bacterial and fungal strains with different degrees, methanol, and ethyl acetate extracts of *S. marianum* showed the broad spectrum against the tested bacteria. The pathogen *E. coli* was the most sensitive bacteria (25 mm), while *A. fumigatus* and *A. niger* were the most sensitive fungi (25 and 27 mm) in case of methanol extract.

The phytotoxicity of *S. marianum* extracts was increased significantly with the increasing extract concentrations. At 25 g.l⁻¹ and 20 g.l⁻¹ the germination of *C. murale* was reached maximum inhibition (92.45% and 76.84, respectively). On the other hand, in case of *C. murale* radicle growth, the higher concentration (25 g.l⁻¹) was strongly inhibited (97.23%) the *C. murale*. Similarity, the plant extract significantly reduced the plumule length of *C. murale*. The highest concentrations (25 g.l⁻¹) of extract showed strong inhibition (87.29%) of plumule growth. Finally, this study showed that *S. marianum* extracts can be used as a natural antioxidant and antimicrobial agents in pharmaceutical as well as used in bio-control of weeds.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- World Health Organization (WHO). Who guidelines on safety monitoring of herbal medicines in pharmacovigilance systems. Geneva, Switzerland: World Health Organization; 2004.
- Chintamunnee V, Mahomoodally MF. Herbal medicine commonly used against infectious diseases in the tropical island of Mauritius. *J. Herbal Med.* 2012;113–125.
- Nunkoo H, and Mahomoodally MF. Ethnopharmacological survey of native remedies commonly used against infectious diseases in the tropical island of Mauritius, *J. Ethnopharmacol.* 2012; 143(2):548–564.
- Gurib-Fakim A, Mahomoodally MF. African flora as potential sources of medicinal plants: Towards the chemotherapy of major parasitic and other infectious diseases- a review. *Jordan J. Biol. Sci.* 2013;6:77–84.
- Kouri G, Bardouki H. Extraction and analysis of antioxidant components from *Origanum dictamnus*. *Innov. Food Sci. & Emerg. Technol.* 2007;8:155-162.
- Panchawat S, Rathore KS, Sisodia SS. A review on herbal antioxidants. *Int. J. Pharm. Tech. Res.* 2010;2(1):232-239.
- Nasir YJ. Flora of Pakistan. Asteraceae. National herbarium Pakistan agriculture research council. Islamabad. 1990;192: 59-117.
- Burgess CA. *Silybum marianum* (Milk Thistle). *Journal of Pharmaceutical Society, Wincons.* 2003;38-40.
- Rajiha A. The therapeutic effect of *Silybum marianum* the lead acetate induced – reproductive toxicity in both gender laboratory rats. *J. I Sci. & Med.* 2012;5(1): 144–15.
- Sayyah M, Boostania H, Paksereshta S, Malayeria A. Comparison of *Silybum marianum* (L.). Gaertn. With fluoxetine in the treatment of obsessive-compulsive disorder. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry.* 2011;35(4):887-895.
- Boulos L. Flora of Egypt. Al-Hadara Pub., Cairo, Egypt. 2002;3.
- Mehraban F, Nasim OT, Fereshteh J. Antidermatophyte activities *Eucalyptus comaldulensis* in comparism with Griscofulvin. *Iran. J. Pharmacol. Ther.* 2005;4:80-83.
- Harborne JB, *Phytochemical methods*, Chapman and Hall, Ltd., London. 1973; 49-188.
- Sadasivam S, Manickam A. *Biochemical methods*, 3rd Ed., New Age International Limited, New Delhi; 2008.
- Boham BA, Kocipai-Abyazan R. Flavonoids and condensed tannin from leaves of Hawaiian *Vccinium vaticulatum* and *V. calycinium*, *Pacif. Sci.* 1994;48: 458-463.
- Van-Buren JP, Robinson WB, Formation of complexes between protein and tannic acid. *J. Agric. Food Chem.* 1969;17:772-777.
- Obadoni BO, Ochuko PO. Phytochemical studies and comparative efficacy of the crude extracts of some homeostatic plants in edo and delta states of Nigeria. *Global J. Pure Appl. Sci.* 2001;8:203-208.
- Miguel MG. Antioxidant activity of medicinal and aromatic plants. *Flavou. Frag. J.* 2010;25:291-312.

19. Cappuccino JG, Sherman N. Microbiology: A laboratory manual. Pearson/Benjamin Cummings, San Francisco; 2008.
20. Sampietro DA, Catalan CAN, Vattuone, MA. Isolation, identification and characterization of allelochemicals natural products. Science Publishers, Enfield, NH, USA; 2009.
21. Rice CA, Miller NJ, Bolwell PG, Bramley PM, Pridham JB. The relative antioxidant activities of plant derived polyphenolic flavonoids. Free Radic. Res. 1995;22(4): 375–383.
22. Shah SMM, Khan FA, Shah SMH, Chishti KA, Saifur SM, Pirezada S, Khan MA, Farid A. Evaluation of phytochemicals and antimicrobial activity of white and blue capitulum and whole plant of *Silybum marianum*. World Appl. Sci. J. 2011;12(8): 1139-1144.
23. Salem N, Dhifi W, Graya A, Mnafeq F, Gharbi S, Khammassi S, Marzouk B, Limam F, Chekir Ben Chaouacha R. Antioxydant activity of *Silybum marianum* and *Ajugaiva* natural dyes. Int. J. Cont. Ene. Elec. Eng. 2016;3(2):6-12.
24. El-Amier YA, Abdelghan AM, Abed Zaid A. Green synthesis and antimicrobial activity of *Senecio glaucus* - mediated silver nanoparticles. Res. J. Pharm. Biol. Chem. Sci. 2014;5:631-642.
25. El-Amier YA, Al-hadithy ON, Abdullah TJ. Antioxidant and antimicrobial activity of different extracts obtained from aerial parts of *Urospermum picroides* (L.) F.W. from Egypt, J. Adv. Chem. Sci. 2016;2(3):299-301.
26. El-Amier YA, Abdullah TJ. Evaluation of nutritional value for four kinds of wild plants in Northern Sector of Nile Delta, Egypt. O. J. App. S. 2015;5:393-402.
27. Dayanne R, Saulo R, Maria F, Aline A. In vitro antimicrobial and modulatory activity of the natural products silymarin and silibinin. BioMed Res. Int. 2015;Article ID. 292797,7.
28. Izzo AA, Di Carlo G, Biscardi D, De Fusco R, Mascolo N, Borrelli F, Capasso F, Fasulo MP, Autore G. Biological screening of italian medicinal plants for antibacterial activity. Phytotherapy Research. 1995;9: 281-286.
29. Ramdani M, Lograd TA, Chalard P, Figueredo G, Laidoudi H, Ounough AI. Chemical composition, antimicrobial activity and chromosome number of *Urospermum dalechampii* from Algeria, Scholars Acad. J. Pharm. 2014;3:477-482.
30. Michelin D, Moreschi P, Lima A, Nascimento G, Paganelli M, Chaud M. Evaluation of the antimicrobial activity of vegetal extracts. Braz. J. Pharmacog. 2005;15(4):316-320.
31. El-Amier YA, Abbas MA, Dawood SH. Phytotoxic effect of plant extracts from Asteraceae on germination and growth of *Echinochloa crus-galli*. Int. J. Dev. Res. 2015;5(7):4926-4931.
32. El-Shora HM, El-Amier YA, Awad MH. Antimicrobial activity and allelopathic potential of *Zygophyllum coccineum* L. on *Chenopodium album* L. Br. J. Appl. Sci. Technol. 2016;15(5):1-10.
33. Noor M, Salem U, Khan MA. Allelopathic effects of *Prosopis juliflora* Swartz. J. Arid Environ. 1995;31:83-90.
34. Vaid S, Batish DR, Singh H, Kohli R. Phytotoxic effect of eugenol towards two weedy species. The Bioscan. 2010;5:339–341.
35. Gniazdowska A, Bogatek R. Allelopathic interactions between plants. Multi site action of allelochemicals. Acta Physiol Plant. 2005;27(3):395-407.
36. Elisante F, Tarimo MT, Ndakidemi PA. Allelopathic effect of seed and leaf aqueous extracts of *Datura stramonium* on leaf chlorophyll content, shoot and root elongation of *Cenchrus ciliaris* and *Neonotonia wightii*. Am. J. Plant Sci. 2013; 04(12):2332-2339.

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